

In Vivo Effect of Diosmin on Carrageenan and CCl₄-Induced Lipid Peroxidation in Rat Liver Microsomes

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ABSTRACT: The aim of this study was to compare the protective effect of a flavonoid, the 3',5,7-trihydroxy-4'-methoxyflavone 7-rutinoside or diosmin, on liver microsomal lipid peroxidation induced in rats by either carbon tetrachloride or carrageenan. Thirty rats were divided into five groups. Group 1 received no chemical product and was considered as control. Groups 2 and 3 received either an intraperitoneal injection of carrageenan or carbon tetrachloride 48 or 24 hours before killing, respectively. Groups 4 and 5 were treated first with an intraperitoneal injection of diosmin and then by carrageenan (group 4) or carbon tetrachloride (group 5) 48 or 24 hours before killing, respectively. The lipoperoxidant effect of carrageenan and carbon tetrachloride was demonstrated by both significant decreases in polyunsaturated fatty acids, principally 20:4 ($n = 6$) ($p < 0.05$) and of vitamin A ($p < 0.05$) in groups 2 and 3. With diosmin treatment, only thiobarbituric acid reactive substances significantly decreased in group 4, whereas vitamin A level increased. These results could suggest that the effect of diosmin differs with the choice of chemical product used; it seems a better antioxidant against products inducing inflammation. © 1996 John Wiley & Sons, Inc.

KEY WORDS: Carbon Tetrachloride, Carrageenan, Diosmin, Lipid Peroxidation, Liver.

INTRODUCTION

Lipid peroxidation (LPO) has been identified as a deleterious process involved in many pathological disorders such as liver injury (1). It was initiated by an excessive production of reactive oxygen species (ROS) that reacted with membrane lipids. The oxidative degradation of polyunsaturated fatty acids (PUFA) leading to damage of biological membranes is one of the major effects of free radical reactions (2). In physiological conditions, LPO occurs, but many enzymatic systems or antioxidant nutrients protect the biological

membranes. LPO can be detected by measuring the breakdown products of lipid peroxides such as malondialdehyde (MDA) by the simple thiobarbituric acid (TBA) test, as thiobarbituric acid reactive substances (TBARS), which correlates well with other methods for measuring LPO (5).

Carbon tetrachloride (CCl₄) is a hepatotoxic agent that is known to stimulate LPO in rat liver microsomes and that produces also disturbances to other tissues (6). It has been effectively demonstrated (7) that most LPO in the liver of CCl₄-intoxicated rats occurs in the endoplasmic reticulum where most cytochrome P450 is located. The toxicity of CCl₄ is depending upon its metabolic activation to trichloromethyl free radical CCl₃· that can function as a peroxidation chain initiator. CCl₃· combines with O₂ to yield the more reactive trichloromethylperoxy radical CCl₃O₂· (8) that covalently binds to cellular components or sparks a LPO process (9). Some researchers indicated that the damage produced (10) will depend on the ratio between trichloromethyl and trichloromethylperoxy radicals. However, a direct CCl₄ action detected by enzyme leakage has been proposed by Berger et al. (11).

Carrageenan, a long-chain-sulfated polysaccharide, is known to induce inflammation and a lipoperoxidant effect in rats (12). An increased release of many factors like histamin, serotonin, prostaglandins, and leukotrienes has also been observed in the inflammatory tissues (13).

Diosmin, the 3',5,7-trihydroxy-4'-methoxyflavone 7-rutinoside, is one of the flavonoid family that shows antioxidant or free radical scavenger properties. It may reduce in rats the paw oedema induced by doxorubicin (14) and the inflammatory granuloma, affecting the synthesis of prostaglandins and thromboxane (15). This flavonoid protects isolated human lipoproteins from in vitro peroxidation (16) and is extensively metabolized by the liver (17).

In a previous study, we have demonstrated that treatment of rats with diosmin before an injection of carrageenan or CCl₄ decreased in serum the acute inflammatory reaction and the lipoperoxidant effect (12).

Received December 18, 1995.

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The present study was designed to investigate *in vivo* the direct influence of diosmin on rat liver microsomes after carrageenan or CCl_4 intoxication and to find out whether diosmin treatment may compensate for the damage formed in the presence of a high level of peroxides.

MATERIALS AND METHODS

Animals and Treatments

Thirty weanling male Wistar rats weighing 50–60 g were housed individually in a room with a temperature of 20°C. They were fed on commercial food pellets AO3 (UAR, Epinay sur Orge, France). Food and water were provided *ad libitum*. Rats were assigned to the basal diet for 12 weeks. Then they were weighed (400–420 g) and divided into five groups. Group 1 received no treatment before killing and was considered as control. Group 2 received an intraperitoneal injection of 2.5 mL of 2% suspension of carrageenan in 0.15 M NaCl (corresponding to 50 mg carrageenan) per kg body weight, 48 hours before killing. Group 3 received an intraperitoneal injection of 1.25 mL of a 1/4 (V/V) mixture of CCl_4 and vaseline oil (corresponding to 0.31 mL CCl_4) per kg body weight, 24 hours before killing. Groups 4 and 5 were pretreated with an intraperitoneal injection of 1.66 mL of a 90/1 (W/V) suspension of diosmin in 0.15 M NaCl (corresponding to 150 mg diosmin) per kg body weight and per week during the last 8 weeks, which preceded injections of carrageenan (group 4) or CCl_4 (group 5). Food consumption and body weights were measured weekly.

Tissue Preparation

After an overnight fast of 12 hours, rats were killed by decapitation. The livers were quickly excised; rinsed in saline; and then dried, weighed, and stored at -80°C until analyzed. Microsomal suspension was prepared as described previously (18). In brief, the homogenized rat liver (1 g wet weight/3 mL of 10 mM Tris-HCl, 150 mM KCl, 0.1 mM EDTA buffer, pH 7.4) was centrifuged initially at $9,000 \times g$ for 15 minutes at 4°C , and the supernatant was recentrifuged at $105,000 \times g$ for 1 hour at 4°C . The resulting pellet was suspended in 10 mM Tris-HCl, 150 mM KCl buffer, pH 7.4, so as to obtain a concentration of 5–10 mg protein/mL and was used immediately.

Analytical Procedure

Vitamins A and E and retinylpalmitate were quantitated by high-performance liquid chromatography

(HPLC) using a method previously described (19). The results were expressed as pmol/mg of liver microsomal protein.

LPO was evaluated as TBARS, as described by Recknagel et al. (20). Tetraethoxypropane was used as standard. TBARS were spectrophotometrically determined at 532 nm, and the LPO level was expressed as nmol of MDA equivalents/mg of liver microsomal protein. Protein was estimated by the method of Lowry et al. (21) using bovine serum albumin as standard.

Fatty acid composition was determined from 2 mL of a lipid extract after transformation in isopropyl esters (22). Peak identification was made by comparison with reference fatty acids (Sigma, St. Louis, MO). Quantification of each fatty acid was expressed as the percentage of the total extract.

Statistical Analysis

Data are presented as means \pm SD. Statistical analyses were done using a statistical software package (Stat-View). For comparison between paired data, the Mann Whitney test was chosen because of the sample size. A *p* value < 0.05 was considered significant.

RESULTS

At the end of the experimental period, no significant differences in body and liver weights were observed among the five groups.

As compared to control group 1, microsomal vitamin A (Table 1) was significantly decreased after carrageenan or CCl_4 treatment in groups 2 and 3; with diosmin, the decrease was still significant in groups 4 and 5. Retinylpalmitate was only significantly decreased in CCl_4 groups 3 and 5, as compared with group 1. A slight increase was observed between groups 2 and 4 and between groups 3 and 5. Unlike vitamin A, vitamin E levels were similar in all studied groups.

Levels of microsomal TBARS (Table 2) were significantly increased in groups 2 through 4 compared with control group 1, indicating the occurrence of LPO. However, the TBARS amount was significantly decreased in group 4 as compared with group 2, while no significant difference was observed between groups 3 and 5. Comparative spectra of TBARS formed during peroxidation (Figure 1) showed a peak at 532 nm in all cases and a slight absorption at 460 nm in CCl_4 groups 3 and 5.

Total PUFA percentages (Table 2) decreased significantly in groups 2 through 4. Fatty acid composition of experimental groups (Table 3) was determined after

TABLE 1. Vitamin A and E Concentrations^a in Liver Microsomes

Measurement	Groups				
	1 Control ^b	2 Carrageenan	3 CCl ₄	4 Diosmin Plus Carrageenan	5 Diosmin Plus CCl ₄
Vitamin A	366 ± 77 ^c	183 ± 33*	100 ± 16*	207 ± 45*	103 ± 32*
Retinyl palmitate ^d	245 ± 58	160 ± 36	129 ± 19*	197 ± 40	142 ± 17*
Vitamin E	199 ± 51	165 ± 34	146 ± 28	168 ± 27	136 ± 19

^aVitamins A and E are expressed as pmol/mg microsomal protein.

^bControl group received no treatment before killing.

^cValues are means ± SD for six animals per group. Significance level for each group compared with control group 1 was studied by the Mann Whitney test (**p* < 0.05).

^dRetinylpalmitate is expressed as pmol/mg microsomal protein.

TABLE 2. Lipid Peroxide (TBARS) Values^a and Polyunsaturated Fatty Acids (PUFA) Percentages^b in Liver Microsomes

Measurement	Groups				
	1 Control	2 Carrageenan	3 CCl ₄	4 Diosmin Plus Carrageenan	5 Diosmin Plus CCl ₄
TBARS	108 ± 50 ^c	670 ± 131*	190 ± 57*	284 ± 56*	170 ± 44
PUFA	50 ± 1	41 ± 1*	45 ± 1*	47 ± 1*	49 ± 1

^aTBARS values are expressed as nmol/mg microsomal protein.

^bPUFA are expressed as area % of total fatty acids.

^cValues are means ± SD for six animals per group. Significance level for each group compared with control group 1 was studied by the Mann Whitney test (**p* < 0.05).

an overnight fast of 12 hours. The most abundant microsomal PUFA of control rats were arachidonic acid,

20:4 (*n* = 6), 21.73 ± 2.06%; linoleic acid, 18:2 (*n* = 6), 16.53 ± 1.36%; and docosahexaenoic acid, 22:6 (*n* = 3), 7.84 ± 0.71%. In group 2, we observed a significant decrease in 20:4 (*n* = 6) and in 22:6 (*n* = 3) mainly involved in LPO. In group 3, only 20:4 (*n* = 6) significantly decreased. This last PUFA was also depressed in groups 4 and 5. However, diosmin treatment led to an increase in PUFA percentages, higher in group 4 than in group 5, compared with groups 2 and 3, respectively.

DISCUSSION

This study investigated the *in vivo* effect of a flavonoid, the diosmin, on the peroxidative process induced by chemical products, such as carrageenan and CCl₄ in rat liver microsomes.

At the end of the experimental period, body and liver weights of the different groups did not differ one from another, indicating that the nutritional requirements of rats were not altered by the different treatments. However, in serum, the inflammatory effect of carrageenan was demonstrated by elevated levels of *a*₁, *a*₂, and *β* globulins and the hepatotoxicity of CCl₄ proved by the increase of *a*₁ globulin (data not shown).

Microsomal vitamin A levels significantly decreased, more in group 3 than in group 2 versus group 1. Such a decrease in hepatic vitamin A storage was

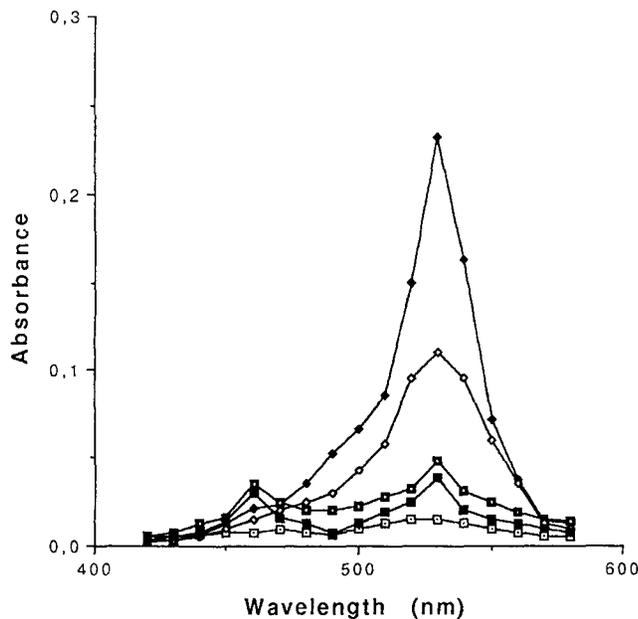


FIGURE 1. Dependence of absorption on wavelength. The absorption was measured in rat liver microsomal suspension after TBA reaction. Reaction conditions are described in Materials and Methods. Symbols used are as follows: □- control (1); -◆-, carrageenan (2); -■-, CCl₄ (3); -◇-, diosmin plus carrageenan (4); -■-, diosmin plus CCl₄ (5).

TABLE 3. Fatty Acid Composition^a of Experimental Groups

Fatty Acid	Groups				
	1 Control	2 Carrageenan	3 CCl ₄	4 Diosmin Plus Carrageenan	5 Diosmin Plus CCl ₄
16:0	21 ± 2 ^b	24 ± 2	23 ± 2	22 ± 2	22 ± 2
16:1 (n - 7)	<2	<2	<2	<2	<2
18:0	20 ± 2	26 ± 2	23 ± 2	23 ± 2	20 ± 1
18:1 (n - 9)	7 ± 1	7 ± 1	8 ± 1	7 ± 1	8 ± 1
18:2 (n - 6)	17 ± 1	16 ± 1	17 ± 2	19 ± 2	19 ± 2
20:4 (n - 6)	22 ± 2	16 ± 1*	16 ± 1*	17 ± 2*	18 ± 2*
20:5 (n - 3)	<2	<2	<2	<2	<2
22:5 (n - 3)	<2	<2	<2	<2	<2
22:6 (n - 3)	8 ± 1	6 ± 1*	8 ± 1	7 ± 1	8 ± 1

^aFatty acids are expressed as area % of total fatty acids. Fatty acid composition was determined after an overnight fast of 12 h.

^bValues are means ± SD for six animals per group. Significance level for each group compared with control group 1 was studied by the Mann Whitney test (**p* < 0.05).

also observed after chronic ethanol consumption (23). It is known that the major site of CCl₄ activation within the liver is the endoplasmic reticulum where most cytochrome P450 is located (7). Since an increase in cytochrome P450 has been reported in rats treated with CCl₄ (23) or with phenobarbital (24), its contribution on the observed decrease in vitamin A level can be suggested. Oxidation of vitamin A and increased catabolism of retinoic acid (25) have also been postulated to explain the vitamin A decrease. The low vitamin A concentration observed with carrageenan can be due to its utilization in the synthesis of glycoproteins (26) and also to its consumption as antioxidant like ascorbate, SH groups, and urate (27). Diosmin treatment led to a slight increase in vitamin A level in groups 4 and 5 (13% and 3%, respectively) compared with groups 2 and 3. Thus the interaction between CCl₄ and vitamin A appeared drastic for the microsomal membrane and led to vitamin A degradation in the liver, as observed after ethanol administration (28). Similarly, retinylpalmitate, the major tissue storage form of liver vitamin A, decreased significantly after CCl₄ intoxication; with diosmin treatment, the decrease was less marked (group 5). In the carrageenan group, retinylpalmitate was not significantly depressed, indicating the lower effect of carrageenan on vitamin A metabolism. Although it is widely accepted that vitamin E acts as an antioxidant and protects biological membranes against free radical damage, microsomal vitamin E level remained quite unchanged in the carrageenan and CCl₄ groups. Miyazawa *et al.* (29) found a relatively large amount of vitamin E in the liver, 24 hours after the CCl₄ dose, as a consequence of antioxidative metabolic adaptation in the rat body. So regeneration of vitamin E (30) by a synergistic action in vivo between vitamins E and C can explain the similar vitamin E level observed especially in carrageenan groups 2 and 4.

Among the markers of LPO, TBARS were mainly increased in the carrageenan group since carrageenan

induced an extensive inflammatory process. The CCl₄ group exhibited a lower but significant increase in TBARS production, as reported in *in vitro* (9, 31) as well as in *in vivo* experiments (32, 33). An explanation of the slight increase observed in group 3 can be that lipid dienyl radicals generated after CCl₄ injection in part decompose to form alkanes that do not undergo significant oxidative degradation to produce MDA (34). The marked decrease in TBARS values between groups 2 and 4 shows the protective antioxidant role of diosmin against the carrageenan inflammatory process. As shown in Figure 1, although MDA is known to be the main aldehydic peroxidation product to have an absorbance maximum at 532 nm after TBA reaction (35), spectra of TBARS in groups 3 and 5 showed another peak at 460 nm, indicating the presence of interfering aldehydic compounds inducing in a slight overestimation of LPO, as observed by others in erythrocyte hemolysate (36). This phenomenon can explain that no significant difference in TBARS values was observed between groups 3 and 5.

Since TBARS are not the exact reflect of LPO (37), determination of PUFA that are the direct targets of ROS seems to be a better marker. A significant decrease in PUFA percentages takes place both in groups 2 and 3 versus control group 1. Peroxidative degradation of unsaturated fatty acids can greatly alter the function of cell membranes (38). The more unsaturated fatty acids are most susceptible to LPO. 20:4 (n - 6) and 22:6 (n - 3) are the major fatty acids to be destroyed during CCl₄-stimulated LPO in liver microsomes (8); they are significantly decreased in our experiments, especially in group 2. Only 20:4 (n - 6) decreased in group 3. With diosmin treatment, the PUFA level increased in groups 4 and 5, compared with groups 2 and 3, respectively.

So the decreased LPO observed in rats treated with diosmin compared with untreated rats may be attrib-

uted to the antioxidant effect of diosmin. However, the difference in the response of the liver microsomes to diosmin correlates well with their susceptibility to carrageenan or CCl_4 -induced LPO.

Our results suggest that diosmin can be used to maintain sufficient protection against liver oxidative damage, especially in the carrageenan inflammatory process. Its antioxidant efficiency may be influenced by several factors, including the rate of radical production and the type of radical initiator. Further studies should be conducted to investigate the biochemical sequences preceding the pathological manifestations of carrageenan and CCl_4 in liver cell membranes.

REFERENCES

- G. Poli, M. U. Dianzani, K. H. Cheeseman, T. F. Slater, J. Lang, and H. Esterbauer (1985). Separation and characterization of the aldehydic products of lipid peroxidation stimulated by carbon tetrachloride or ADP-iron in isolated rat hepatocytes and rat liver microsomal suspensions. *Biochem. J.*, **227**, 629–638.
- A. L. Tappel (1973). Lipid peroxidation damage to cell components. *Fed. Proc.*, **32**, 1870–1874.
- H. W. Leung, M. J. Vang, and R. D. Mavis (1981). The cooperative interaction between vitamin E and vitamin C in suppression of peroxidation of membrane phospholipid. *Biochim. Biophys. Acta*, **664**, 266–272.
- M. E. Murphy and J. P. Kehrer (1989). Lipid peroxidation inhibitory factors in liver and muscle of rat, mouse, and chicken. *Arch. Biochem. Biophys.*, **268**, 585–593.
- Y. Singh, G. L. Hall, and M. G. Miller (1992). Species differences in membrane susceptibility to lipid peroxidation. *J. Biochem. Toxicol.*, **7**, 95–105.
- C. Benedetto, M. U. Dianzani, M. Ahmed, K. Cheeseman, C. Connely, and T. F. Slater (1981). Activation of carbon tetrachloride and distribution of NADPH-cytochrome c reductase, cytochrome P450 and other microsomal enzyme activities in rat tissues. *Biochim. Biophys. Acta*, **677**, 363–372.
- G. D. Castro, M. I. Diaz Gomez, and J. A. Castro (1990). Biotransformation of carbon tetrachloride and lipid peroxidation promotion by liver nuclear preparations from different animal species. *Cancer Lett.*, **53**, 9–15.
- R. N. Le Page, K. H. Cheeseman, N. Osman, and T. F. Slater (1988). Lipid peroxidation in purified plasma membrane fractions of rat liver in relation to the hepatotoxicity of carbon tetrachloride. *Cell. Biochem. Func.*, **6**, 87–99.
- S. L. Fanelli and J. A. Castro (1993). Carbon tetrachloride promoted malondialdehyde formation in liver microsomal and nuclear preparations from Sprague Dawley or Osborne Mendel male rats. *Res. Commun. Chem. Pathol. Pharmacol.*, **82**, 233–236.
- E. A. Lissi and H. Rosenbluth (1993). Disruption effects of carbon tetrachloride on rat liver microsomes. *J. Photochem. Photobiol. B: Biol.*, **17**, 33–40.
- M. Berger, H. Rhatt, B. Combes, and B. Estbrook (1986). Tetrachloride induced toxicity in isolated hepatocytes. The importance of direct solvent injury. *Hepatology*, **6**, 36–45.
- M. Freneix-Clerc, M. F. Dumon, M. A. Carbonneau, M. J. Thomas, E. Peuchant, L. Dubourg, A. M. Melin, A. Perromat, and M. Clerc (1994). Etude *in vivo* de l'effet antilipoperoxidant du 7 rutinoside de la 3',5,7-trihydroxy-4'-methoxyflavone. *Ann. Biol. Clin.*, **52**, 171–177.
- C. G. Van Arman, A. Begany, L. M. Miller, and H. H. Pless (1965). Some details of the inflammation caused by yeast and carrageenin. *J. Pharmacol. Exp. Ther.*, **150**, 328–334.
- E. Sauvaire, A. Michel, J. J. Serrano, and M. Reynier (1989). Effects of flavonoids (Diosmin and Silymarin) in rat adriamycin-induced paw edema. Third Intersci. World Conf. on Inflammation (Abstract), Monte Carlo, pp. 332.
- P. Damon, O. Flandre, F. Michel, L. Perdix, C. Labrid, and A. Crastes de Paulet (1987). Effect of chronic treatment with a purified granuloma in the rat. Study of prostaglandin E_2 et $F_{2\alpha}$ and thromboxane B_2 release and histological changes. *Arzneim Forsch.*, **37**, 1149–1153.
- M. F. Dumon, M. Freneix-Clerc, M. A. Carbonneau, M. J. Thomas, E. Peuchant, A. Perromat, and M. Clerc (1994). Mise en évidence de l'effet antilipoperoxidant du 7 rutinoside de la 3',5,7-trihydroxy-4'-methoxyflavone: étude *in vitro*. *Ann. Biol. Clin.*, **52**, 265–270.
- R. Perego, P. Beccaglia, M. Angelini, P. Villa, and D. Cova (1993). Pharmacokinetic studies of diosmin and diosmetin in perfused rat liver. *Xenobiotica*, **23**, 1345–1352.
- A. M. Melin, M. A. Carbonneau, M. J. Maviel, A. Perromat, and M. Clerc (1990). Free radical inhibitor effect of retinol after carbon tetrachloride intoxication in the rat. *Food Add. Contam.*, **7**, S182–S187.
- A. M. Melin, M. A. Carbonneau, M. J. Thomas, M. J. Maviel, A. Perromat, and M. Clerc (1992). Relationship between dietary retinol and α -tocopherol and lipid peroxidation in rat liver cytosol. *Food Add. Contam.*, **9**, 1–9.
- R. O. Recknagel, E. A. Glende, R. L. Waller, and K. Lowrey (1982). Lipid peroxidation biochemistry, measurement and significance in liver cell injury. In *Toxicology of the Liver*, G. L. Plaa and W. R. Hewitt, eds., pp. 213–241, Raven Press, New York.
- O. H. Lowry, N. J. Rosebrough, A. L. Farr, and R. J. Randall (1951). Protein measurement with the Folin phenol reagent. *J. Biol. Chem.*, **193**, 265–275.
- E. Peuchant, R. J. Wolff, C. Salles, and R. Jensen (1989). One-step extraction of human erythrocyte lipids allowing rapid determination of fatty acid composition. *Anal. Biochem.*, **181**, 341–344.
- K. Chapman, M. Prabhudesai, and J. W. Erdman (1992). Effects of ethanol and carbon tetrachloride upon vitamin A status of rats. *Alcohol. Clin. Exp. Res.*, **16**, 764–768.
- M. A. Leo, N. Lowe, and C. S. Lieber (1987). Potentiation of ethanol-reduced hepatic vitamin A depletion by phenobarbital and butylated hydroxy tolerance. *J. Nutr.*, **117**, 70–76.
- M. Sato and C. S. Lieber (1982). Increased metabolism of retinoic acid after chronic ethanol consumption in rat liver microsomes. *Arch. Biochem. Biophys.*, **213**, 557–564.
- G. Wolf, T. C. Kiorpes, S. Masushige, J. B. Schreider, M. J. Smith, and R. S. Anderson (1979). Recent evidence for the participation of vitamin E in glycoprotein synthesis. *Fed. Proc.*, **38**, 2540–2543.
- J. Muntane, P. Puig Parellada, Y. Fernandez, S. Mitjavila, and M. T. Mitjavila (1993). Antioxidant defenses and its modulation by iron in carrageenan-induced inflammation in rat. *Clin. Chim. Acta*, **214**, 185–193.
- M. A. Leo, N. Lowe, and C. S. Lieber (1986). Interactions of drugs and retinol, *Biochem. Pharmacol.*, **25**, 3949–3953.

29. T. Miyazawa, T. Suzuki, K. Fujimoto, and T. Kaneda (1990). Phospholipid hydroperoxid accumulation in liver of rats intoxicated with carbon tetrachloride and its inhibition by dietary α -tocopherol. *J. Biochem.*, **107**, 689–693.
30. J. E. Packer, T. F. Slater, and R. L. Willson (1979). Direct observation of a free radical interaction between vitamin E and vitamin C. *Nature*, **278**, 737–738.
31. G. H. I. Wolfgang, W. J. Donarski, and T. W. Petry (1990). Effects of novel antioxidants on carbon tetrachloride-induced lipid peroxidation and toxicity in precision-cut rat liver slices. *Toxicol. Appl. Pharmacol.*, **106**, 63–70.
32. F. P. Corungiu, M. Lai, and F. Milia (1983). Carbon tetrachloride, bromo trichloromethane and ethanol acute intoxication. *Biochem. J.*, **212**, 625–631.
33. J. W. Allis, T. R. Ward, J. C. Seely, and J. E. Simmons (1990). Assessment of hepatic indicators of subchronic carbon tetrachloride injury and recovery in rats. *Fundam. Appl. Toxicol.*, **15**, 558–570.
34. H. H. Draper, L. Polensek, M. Hadley, and L. G. McGirr (1984). Urinary malondialdehyde as an indicator of lipid peroxidation in the diet and in the tissues. *Lipids*, **19**, 836–843.
35. H. Ohkawa, N. Ohishi, and K. Yagi (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal. Biochem.*, **95**, 351–358.
36. H. S. Gilbert, D. D. Stump, and E. F. Roth, Jr. (1984). A method to correct for errors caused by generation of interfering compounds during erythrocyte lipid peroxidation. *Anal. Biochem.*, **137**, 282–286.
37. E. Bruna, E. Petit, M. Beljean-Leymarie, S. Huynh, and A. Nouvelot (1989). Specific susceptibility of docosahexaenoic acid and eicosapentaenoic acid to peroxidation in aqueous solution. *Lipids*, **24**, 970–975.
38. M. T. Smith, H. Thor, P. Hartzell, and S. Orrenius (1982). The measurement of lipid peroxidation in isolated hepatocytes. *Biochem. Pharmacol.*, **31**, 19–26.