

Stimulatory Effect of a Specific Substance P Antagonist (RPR 100893) of the Human NK₁ Receptor on the Estradiol-Induced LH and FSH Surges in the Ovariectomized Cynomolgus Monkey

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Utilizing a human NK₁ receptor antagonist (RPR 100893), the present *in vivo* study was designed to test the hypothesis that endogenous substance P (SP) modulates the action of 17 β -estradiol in inducing luteinizing hormone (LH) and follicle stimulating hormone (FSH) surges in ovariectomized cynomolgus monkey. Plasma concentrations of LH and FSH as well as NK₁ receptor antagonist and SP were measured during the development of the negative and positive feedback phases which follow a single administration of estradiol benzoate (50 μ g/kg) to long-term ovariectomized monkeys. Daily administration by gastric intubation of 1 mg/kg or 10 mg/kg of the NK₁ receptor antagonist (RPR 100893) leads to detectable levels of the antagonist in the blood of treated animals for at least 6 hr after its administration. These levels are in agreement with the experimentally determined IC₅₀ value of the antagonist.

The most striking finding of this study is that LH and FSH releases are enhanced during the descending arm of the estradiol benzoate-induced LH and FSH surges, which suggests that endogenous SP normally has an inhibitory role during this time. The enhancement of LH release is approximately 50%, regardless of the amount of the NK₁ antagonist used. However, the enhanced FSH release is more important. Furthermore, blockade of the NK₁ receptor with the smaller dose of the antagonist leads to a small, but significant, increase in plasma levels of SP, indicating that blockade of SP receptors leads to an increased release of SP. Collectively, these results further substantiate the link which exists between the ovarian steroid 17 β -estradiol and SP systems. Also, for the first time, these results demonstrate an inhibitory involvement of the human

NK₁ receptor in the 17 β -estradiol-induced pseudo-ovulatory gonadotropin surges in the ovariectomized monkey. *J. Neurosci. Res.* 50:94–103, 1997.

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INTRODUCTION

It has been well documented that the midcycle 17 β -estradiol (E₂) surge is the major event that synchronizes identified and nonidentified interactive mechanisms operating within the hypothalamo-pituitary-complex leading to the midcycle luteinizing hormone (LH) surge (Karsch et al., 1973; Nakai et al., 1978; Knobil and Hotchkiss, 1988). In the ovariectomized (OVX) monkey, a midcycle-like LH surge can be induced by the administration of E₂ benzoate (E₂B), sufficient to provoke circulating concentrations of E₂ exceeding 150 pg/ml for more than 30 hr (Knobil, 1980). It is generally accepted that pulses of GnRH constitute the signal for the tonic pulsatile secretion of LH and follicle stimulating hormone (FSH) during the cycle. However, during the midcycle gonadotropin surge, additional factors, including steroids and various neuropeptides, are implicated.

Previous studies indicate that neuropeptide Y, galanin, neurotensin, δ -sleep-inducing peptide and angiotensin II are involved in the facilitation of the midcycle LH surge in the rat, by an action at the hypothalamic and/or

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anterior pituitary (AP) level (references in Kalra, 1993). On the contrary, substance P (SP), a neurokinin, inhibits the amplitude of the midcycle LH surge in the rat (Battmann et al., 1991). Neurokinins form a family of peptides with neurotransmitter/neuromodulator activities (references in Jessop et al., 1992). These are expressed in neurons throughout the central and peripheral nervous system, and also in endocrine anterior pituitary cells. Substance P, neurokinin A (NKA) and two extended forms of NKA, neuropeptide K (NPK) and neuropeptide γ (NP γ), are encoded by the same preprotachykinin (PPT) gene. The PPT primary transcript is subjected to differential splicing, and the different mRNA isoforms encode different combinations of tachykinins (Krause et al., 1987).

In the rat, a SP-containing hypothalamic neuronal system projects to the median eminence (Makara et al., 1986). Substance P-immunoreactive boutons contact the capillaries of the hypothalamo-pituitary portal system. Within the hypothalamus of the monkey, numerous SP-containing cells have been found in the arcuate nucleus and lateral areas as well as in the periventricular area of the dorsal tuberal region (Ronnekleiv et al., 1985). Estradiol up-regulates hypothalamic SP and NKA content (Akesson, 1994), and increases PPT gene expression (Brown et al., 1990; Priest et al., 1995). Physiological SP and NKA variations in the female hypothalamus during the estrous cycle have also been described (Parnet et al., 1990; Duval et al., 1996a). More recently, variations in levels of substance P-encoding β -, γ -PPT and substance P receptor NK-1 transcripts were documented in the rat hypothalamus throughout the estrous cycle (Gautreau et al., 1997).

The concentrations of SP in the brain, median eminence, arcuate nucleus, medial preoptic nucleus, and AP of rats (Parnet et al., 1990; Antonowicz et al., 1985; De Palatis et al., 1985) appear to be modulated by ovarian steroids. Furthermore, the number of SP-binding sites in the AP varies during the rat estrous cycle (Kerdelhué et al., 1985). Several studies have investigated the role of SP in hypothalamo-pituitary function in monkeys during the induction of a preovulatory-like LH surge following treatment with estradiol benzoate. Substance P delays the occurrence of the E₂B-induced LH surge in OVX monkeys (Jeanjean et al., 1988). In addition, a depletion of AP content of SP is noted in E₂B-treated OVX monkeys (Kerdelhué et al., 1993) by 48 hr post-E₂B, the time of the LH surge. On the other hand, E₂B exposure of OVX monkeys leads to an almost linear increase, up to 48 hr post-E₂B treatment (the time of the E₂-induced LH surge), of the *in vitro* hypothalamic secretion of SP (Kerdelhué et al., 1992b). An acute release of SP from hypothalamus occurred following *in vitro* treatment with progesterone when the hypothalamus had been excised 48

hr post-E₂B treatment (Kerdelhué et al., 1992b). The inhibitory effect of SP is at least exerted at the AP level in the rat (Battmann et al., 1991; Kerdelhué et al., 1978a), monkey (Kerdelhué et al., 1992a) and human (Wormald et al., 1989).

To date, three categories of neurokinin receptors which bind SP to some degree have been documented: NK₁ with primary affinity for SP, NK₂ with primary affinity for NKA and NK₃ with primary affinity for neurokinine B. NK₁ receptors have been characterized on lactotropes and gonadotropes cells at the AP level (Kerdelhué et al., 1985; Larsen et al., 1989, 1992). NK₁ mRNA encodes a seven-transmembrane domains tachykinin receptor, with preferential affinity to SP (Hershey and Krause, 1990). NK₁ is widely expressed in the central and peripheral nervous system (Hershey et al., 1991; Kiyama et al., 1993), and in the anterior pituitary (Winkler et al., 1995).

Recently, a highly potent and specific NK₁ receptor antagonist (RPR 100893) has been synthesized (Tabart and Peyronel, 1994) and characterized (Lee et al., 1994; Cutrer et al., 1995; Fardin et al., 1994a,b; Moussaoui et al., 1994). The NK₁ receptor antagonist RPR 100893 is a potent SP antagonist in models which are predictive of several pathological states, such as asthma, inflammation and pain, migraine and vascular headaches.

Utilizing this NK₁ receptor antagonist, the present *in vivo* study was designed to test the hypothesis that endogenous SP modulates the action of E₂ in inducing LH and FSH surges in primates. The changes in E₂, LH and FSH serum concentrations, during treatment with a specific SP antagonist of the NK₁ receptor (RPR 100893) were quantitated. Plasma levels of this NK₁ receptor antagonist as well as SP, were also measured.

MATERIALS AND METHODS

Fifteen long-term ovariectomized (OVX) adult female cynomolgus monkeys (*Macaca fascicularis*), weighing 3.5 ± 1.6 kg, were used for this study. The monkeys were housed in individual cages and fed a diet of commercial monkey chow (Agways, Elizabeth, NJ). Water was available *ad libitum*; the lighting schedule was 12 hr of light, 12 hr of darkness, with light on from 0600–1800 hr daily; room temperature was maintained at approximately 20°C.

Fifteen primates were randomly divided into three equal groups. At 6 a.m., OVX monkeys received intragastric administrations of 1 mg/kg or 10 mg/kg of RPR 100893 or vehicle (1 ml of polyethylene glycol [PEG 400]) for 5 consecutive days. Six hr after the first administration of the vehicle, or the second doses of the NK₁ antagonist, all 15 OVX monkeys received a subcutaneous injection of estradiol benzoate (E₂B), 50 μ g/kg, in

sesame oil. Blood samples (5 ml) were collected via femoral vein puncture before the administration of the NK₁ antagonist and every 6 hr for 5 days and then every 12 hr for an additional 2 days. Blood samples and treatments were performed under ketamine anesthesia (10 mg/kg, i.m.). Plasma samples were obtained and, after centrifugation, stored frozen at -20°C until assayed by radioimmunoassay (RIA) for 17β-estradiol, LH, FSH and SP.

Radioimmunoassays of 17β-Estradiol, LH, FSH and SP

17β-estradiol. The 17β-estradiol assay was performed using a commercial kit (RSL Inc., Carson, CA).

LH. The LH assay was performed using an antiserum directed against hCG (R₁₃, pool D), a ¹²⁵I-LH preparation from cercopithecus monkeys (WP-XV-117-3239) and a LH preparation from rhesus monkey (LH-RP₁, WP-XW-20). The intra-assay coefficients of variation was less than 9%. The sensitivity of the assay was 1.8 ng/tube.

FSH. Concentrations of monkey FSH were determined using a rabbit antibody to ovine FSH (H-31), preparations of hFSH (NIH-FSH-3) for iodination, and cynomolgus FSH (NICHD-CYN-FSH-RP1) as reference preparation. Two assays were made on all samples. The intra-assay coefficients of variation were less than 8%. The sensitivity of the assay was 1.8 ng/tube.

Substance P. Sep-Pack columns (Waters Associates, Milford, MA) filled with a stationary nonpolar phase (Sep-Pack C18) were used for the preparation of the samples prior to the SP assay, as previously described (Kerdelhué et al., 1992b). Briefly, cartridges were prepared by successive washes with 5 ml ethanol, 5 ml 8 M Urea and 10 ml of bidistilled water before the introduction of 2 ml of plasma on the column. Then the column was washed with 10 ml of H₂O and 10 ml of 4% acetic acid. Peptides retained on the column were eluted with 5 ml of a mixture of 90% ethanol and 4% acetic acid. After a speed vac evaporation (Savant Instruments Inc., Farmingdale, NY) for 12 hr the residuum was reconstituted in 0.5 ml of the RIA buffer and aliquots of 0.1 ml and 0.3 ml were used for the assay. The SP assay was performed as previously described (Cheramy et al., 1978). The intra- and interassay coefficients of variation were 10% and 12% at binding levels of 60%, respectively. The lower limit of detection was 2 pg per tube.

Plasma Substance P Receptor Antagonist (RPR 100893) Assay

The plasma concentration of NK₁ receptor binding compounds (including RPR 100893 and any active metabolite, in proportion to the affinity for a specific

receptor) as determined by radioreceptor assay. The method is based on the competition between the labeled substance P (iodinated ¹²⁵I-Bolton Hunter Substance P, ¹²⁵I-BH-SP) and NK₁ receptor binding compounds present in plasma for binding sites on CHO cell clones selectively expressing human NK₁ receptors. The method involves acetonitrile deproteinization of plasma as pretreatment. A standard curve was established using seven increasing concentrations of RPR 100893 (from 5 to 320 ng/ml). The batch used for bioanalytical measurement is LJP 7194. For this reason, results were expressed as RPR 100893 equivalent. The interassay precision of the method, evaluated from two different concentrations (20 and 100 ng/ml in plasma), shows coefficients of variation lower than 13%. The limit of quantitation was 5 ng RPR 100893 equivalent per ml of plasma using 0.1 ml of sample.

Statistical Analysis

The data were analyzed by a 3 × 25 repeated measures analysis of variance (ANOVA). The 3 represents the levels of the treatment factor, control, 1.0 mg/kg and 10 mg/kg, and the 25 represents the number of repeated measurements from 0 to 174 minutes. Because of the time series structure of the data (the large number of repeated measurements), variance-covariance matrix of the 25 repeated measurements was modeled as first-order autoregressive. This takes into account the fact that measurements closer together in time are more highly correlated than measurements farther apart in time. The ANOVA tested main effects for treatment groups (differences in treatments means pooled over all times), time (trends over time for all groups combined) and the interaction of group and time (differing trends among the treatment groups).

Animal Welfare Assurance

This study was conducted following approval by the Institutional Animal Care and Use Committee of the Eastern Virginia Medical School. This committee conducts its reviews in accord with the NIH *Guide for the Care and Use of Laboratory Animals*, and *Public Health Service Policy on Humane Care and Use of Laboratory Animals*. The facilities of the Division of Animal Resources of the Eastern Virginia Medical School were and are fully accredited by the American Association for the Accreditation of Laboratory Animal Care.

RESULTS

Serum E₂ Concentrations (Fig. 1)

For all groups, there was a significant increase in serum estradiol concentrations from 0 to 6 hr post-E₂B

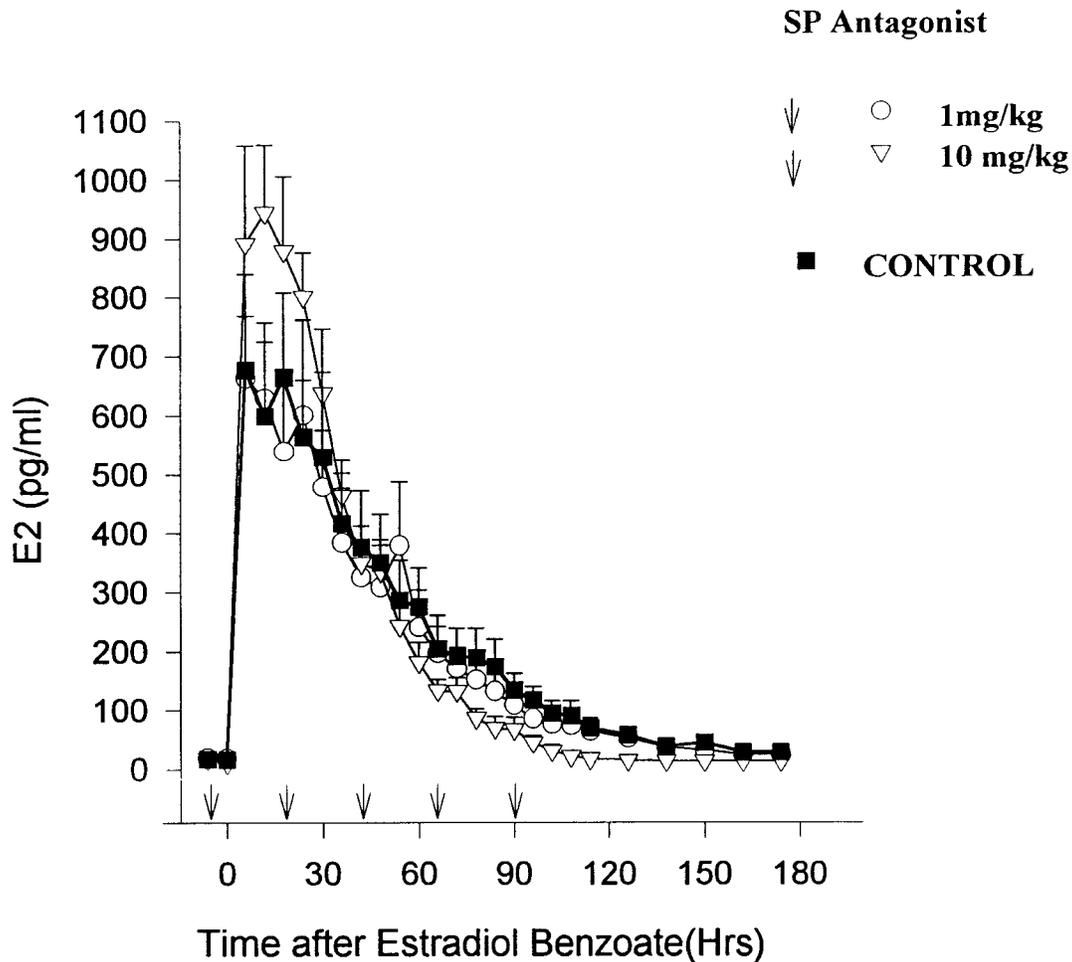


Fig. 1. Pattern of mean (\pm S.E.M.) plasma concentrations of E₂ in blood samples taken up every 6 hr to the time of 174 hr, from OVX cynomolgus monkeys treated with one injection of E₂B (50 μ g/kg) at 0 hr and with five intragastric administrations of 1 mg/kg or 10 mg/kg of RPR 100893 or vehicle, at -6 h, 18, 42, 66, and 90 hr.

treatment, and an exponential decrease after 6 hr ($P < 0.0001$ for time main effect). There was no overall difference between control and NK₁ antagonist-treated animals ($P = 0.08476$ for group main effect) or any difference in the trend between the three groups ($P = 0.7723$ for the group by time interaction).

Serum NK₁ Antagonist Concentrations (Fig. 2)

Maximum levels of NK₁ antagonist were observed 6 hr after each administration of 1 mg/kg or 10 mg/kg of the antagonist. Then levels were between 10 and 60 ng/ml after administration of 1 mg/kg and between 70 and 200 ng/ml after administration of 10 mg/kg of the antagonist.

Serum SP Concentrations (Fig. 3)

There was a significant main effect for the treatment group ($P < .0001$). Overall mean levels for control

monkeys were lower than NK₁ antagonist-treated monkeys. Mean values in control animals were 7.66 ± 0.2 pg/ml as compared to 12.37 ± 0.5 pg/ml (1 mg/kg of NK₁ antagonist) and 9.01 ± 0.3 pg/ml (10 mg/kg of NK₁ antagonist). Values in the 1 mg/kg NK₁ antagonist-treated group were significantly higher than in the control group.

Serum LH Concentrations (Fig. 4)

In control animals, the pattern of mean plasma LH levels followed the expected profile of an E₂B-induced LH surge; castrate levels of LH were initially inhibited by the E₂B treatment, followed by a surge of LH with subsequent reinhibition. Maximum levels occurred by 54 hr post-E₂B treatment. A similar overall pattern of LH concentrations was present in NK₁ antagonist-treated monkeys; maximum levels were observed by 60 hr post-E₂B treatment. Markedly higher serum LH concentrations during the descending arm of the LH surge were

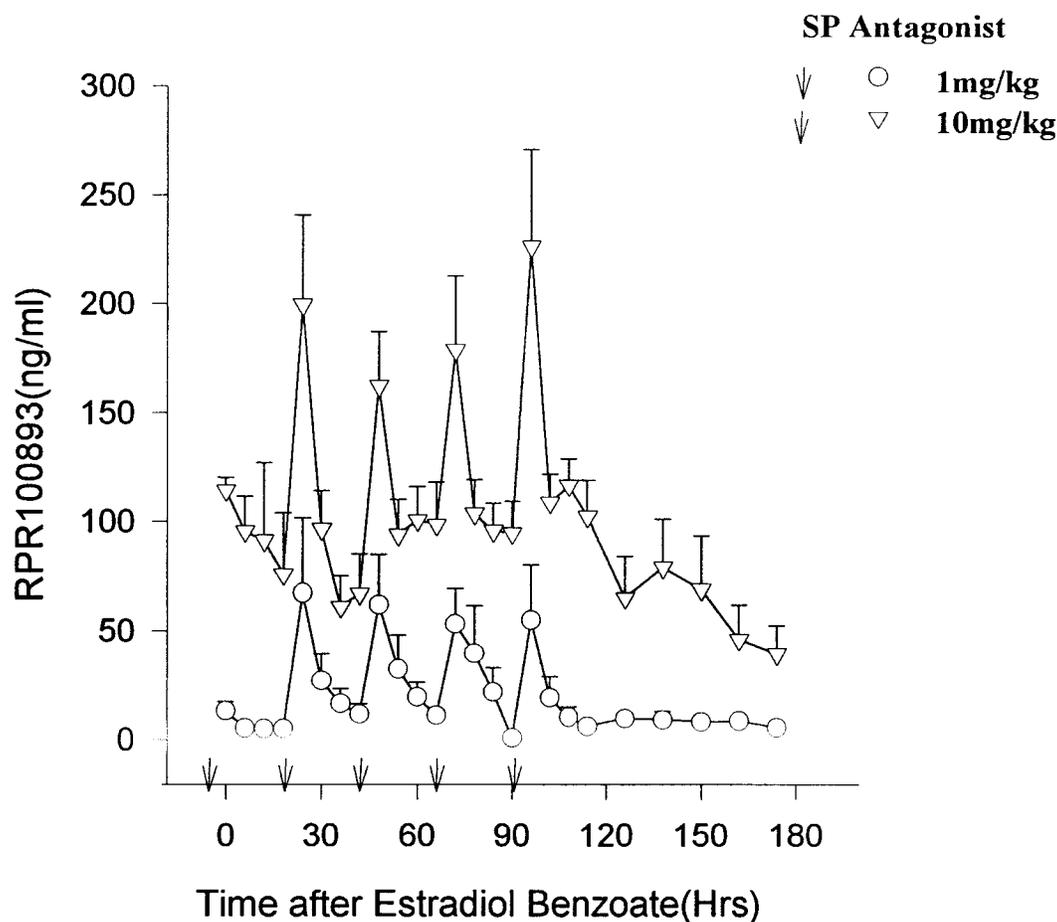


Fig. 2. Pattern of mean (\pm S.E.M.) plasma concentrations of NK₁ antagonist (RPR 100893) in all blood samples taken up every 6 hr to the time of 174 hr, from OVX cynomolgus monkeys treated with one injection of E₂B (50 μ g/kg) at 0 hr and with five intragastric administrations of 1 mg/kg or 10 mg/kg of RPR 100893 or vehicle, at -6, 18, 42, 66, and 90 hr.

present in monkeys treated with the NK₁ antagonist. These increased serum levels of LH persisted for the duration of the study. LH concentrations were 59 ± 22 ng/ml in control and 175 ± 64 and 224 ± 40 ng/ml, after treatments with 1 mg or 10 mg of NK₁ antagonist, respectively, between the 90th and 174th hr of the study. All together, there was a significant main effect for the treatment group ($P = 0.0359$). It can be seen from Figure 4 that controls have the smallest overall mean, primarily because of the values after 60 hr. The overall mean LH of controls was 156.66 ng/ml compared to 275.26 ng/ml for the 1 mg/kg and 235.54 ng/ml for the 10 mg/kg of the NK₁ antagonist.

Serum FSH Concentrations (Fig. 5)

In control monkeys, the pattern of mean plasma FSH followed the expected profile of an E₂B-induced FSH surge. Minimum levels occurred at 42 hr and

maximum levels occurred by 54 hr post-E₂B. In treated animals, the time of maximum levels occurred by 60 hr in animals treated with 1 mg/kg of NK₁ antagonist and by 54 hr in animals treated with 10 mg/kg of the NK₁ antagonist. In addition, the amplitude of the FSH surge was slightly increased in animals treated with 1 mg/kg or 10 mg/kg of NK₁ antagonist. Markedly higher serum FSH concentrations during the descending arm of the FSH surge were present in monkeys treated with the NK₁ antagonist. This increase was still present by 174 hr post-E₂B treatment: Overall mean FSH concentrations were 52 ± 1 ng/ml in control vs. 68 ± 3 ng/ml in 1 mg/kg and 82 ± 10 ng/ml in 10 mg/kg of NK₁ antagonist treated animals ($P < 0.05$).

DISCUSSION

It has been well documented that administration of E₂ to OVX animals induces at the hypothalamo-pituitary

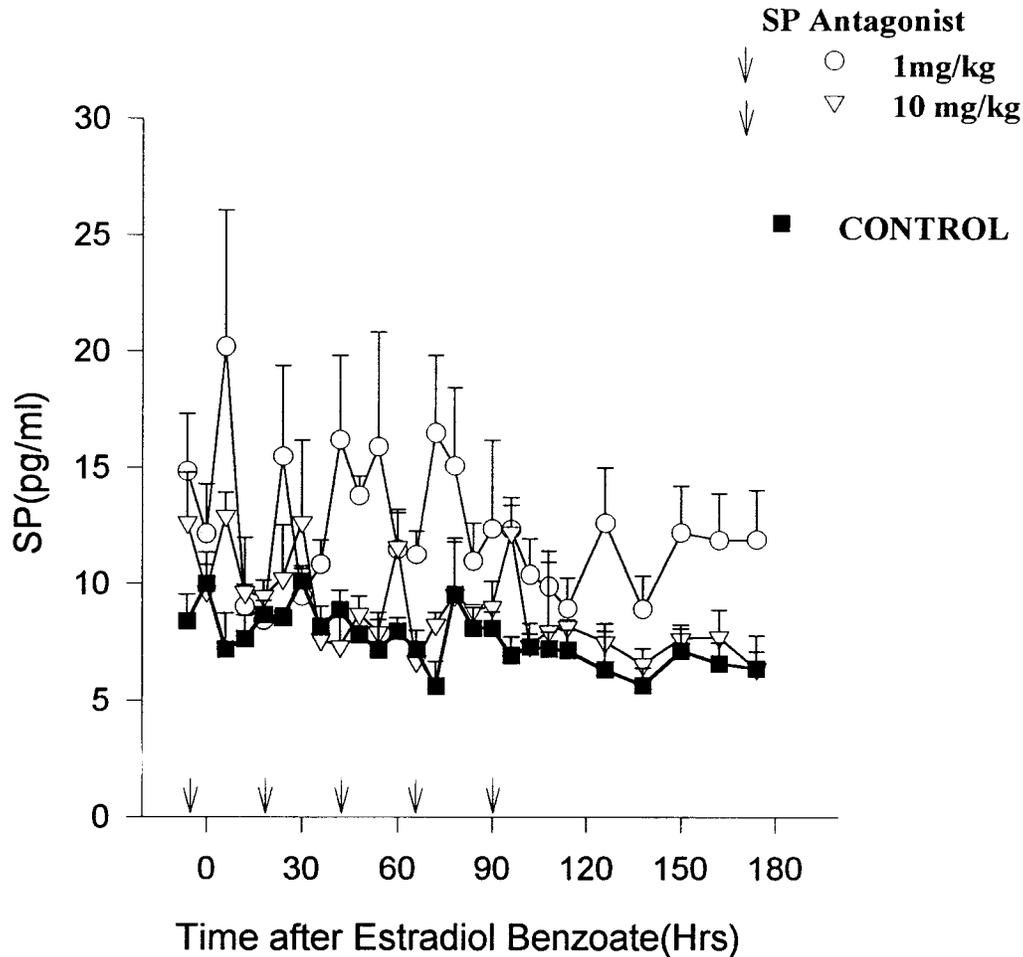


Fig. 3. Pattern of mean (\pm S.E.M.) plasma concentrations of SP in blood samples taken up every 6 hr to the time of 174 hr, from OVX cynomolgus monkeys treated with one injection of E₂B (50 μ g/kg) at 0 hr and with five intragastric administrations of 1 mg/kg or 10 mg/kg of RPR 100893 or vehicle, at -6, 18, 42, 66, and 90 hr.

level all neuroendocrine activations induced by the high level of E₂ which occur at midcycle and are necessary for LH and FSH preovulatory surges (Karsch et al., 1973; Knobil, 1980; Knobil and Hotchkiss, 1988). In this process, progesterone, either from ovarian or adrenal origin, amplifies the stimulatory action of E₂ but is not necessary for the inductive effect of E₂ (Mahesh and Brann, 1992).

The present study demonstrates for the first time an inhibitory involvement of the NK₁ receptor in the post-E₂-induced LH and FSH surges in the OVX monkey. The most striking finding is that LH and FSH concentrations are enhanced in the presence of the NK₁ receptor antagonist during the descending arm of the LH and FSH surge. This finding suggests that SP normally has an inhibitory role during this period of time. The amplitude of the LH surge was not changed but the amplitude of the FSH surge was slightly increased. Although in some

experimental groups (1 and 10 mg/kg for LH and 1 mg/kg for FSH) the peak levels of LH or FSH surge occurred 6 hr later than in controls, as blood collections were made with a 6-hr interval, this difference is not statistically significant. These results are in agreement with previous results which suggested a major secretion of SP from the hypothalamus during the descending arm of an estradiol-induced LH surge (Kerdelhué et al., 1992b). Clearly, there was a clear significant dissociation of the effect of the two different doses of the NK₁ antagonist on FSH release, the higher dose (10 mg/kg) being more efficient on FSH release than on LH release.

A dissociation of LH and FSH release has been previously observed in other physiological or experimental situations such as the immunoneutralization of circulating levels of GnRH (Kerdelhué et al., 1976). In addition, the higher dose (10 mg/kg) down-regulates SP plasma levels to control levels. The reason for the lack of

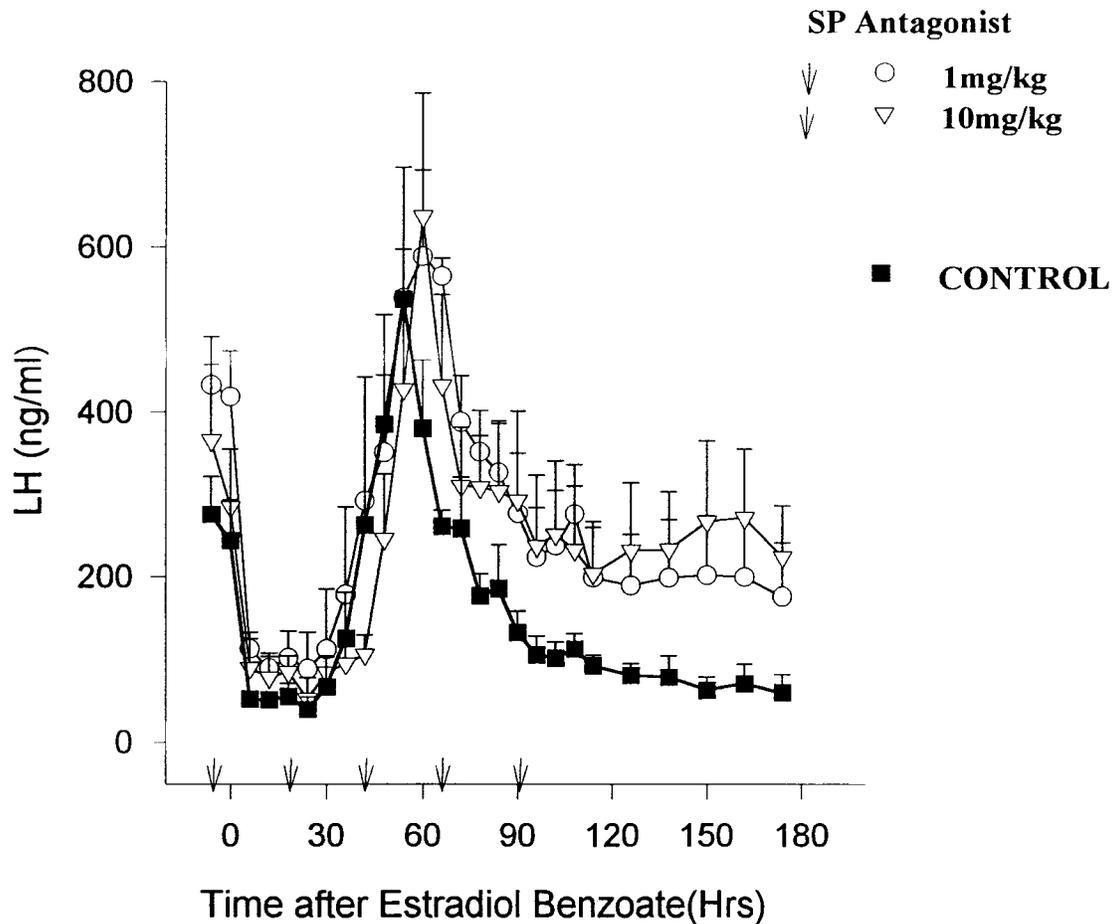


Fig. 4. Pattern of mean (\pm S.E.M.) plasma concentrations of LH in blood samples taken up every 6 hr to the time of 174 hr, from OVX cynomolgus monkeys treated with one injection of E₂B (50 μ g/kg) at 0 hr and with five intragastric administrations of 1 mg/kg or 10 mg/kg of RPR 100893 or vehicle, at -6, 18, 42, 66, and 90 hr.

effect of the NK₁ antagonist on the timing or magnitude of the LH and FSH surge after daily injections of the antagonist is not obvious, since an intracerebroventricular administration of SP delays the onset of the E₂-induced LH surge in the OVX monkey (Jeanjean et al., 1988). In the present study, the stimulatory effect of the antagonist was mainly seen after the peak levels of gonadotropins. It is possible that even though SP has much greater affinity for the NK₁ receptor, it has a weak affinity for the NK₂ and/or NK₃ receptor which can also be involved in LH release during the initial phase of LH release.

In this respect, using binding to various receptors and channels, the RPR 100893 was shown to be highly specific for NK₁ receptor up to 1 μ M. In particular, it does not interact with the NK₂ or the NK₃ receptor nor with opiates receptors (μ , δ or κ) and calcium channels (Fardin et al., 1994a). Therefore, using this NK₁ antagonist, we can assume that the effects observed in the present study are mediated through the NK₁ receptor, although it is

difficult to assess the specificity of the RPR 100893 at the monkey hypothalamic level, since NK₂ receptors were not documented in the brain of various species by direct binding even if the presence of NKA has been documented.

Whether the RPR 100893 exerts its effects through the classical NK₁ receptor and/or a subsite or subtype of the NK₁ receptor sensible to the 6-11 analog of SP (septide) is not known since this compound is able to inhibit to the same extent the effect of SP and septide using in vitro experiments (guinea pig ileon and second messengers production) with a human astrocyte cell line (Fardin et al., 1994a). In fact, plasma levels of the NK₁ receptor antagonist which are in the range of 60 ng/ml (after administration of 1 mg/kg of antagonist) to 200 ng/ml (after administration of 10 mg/kg antagonist) are in agreement with receptor occupancy. A experimentally determined IC₅₀ value (membrane preparation and only 30 min of incubation at 25°C) was 44 nM at the putamen

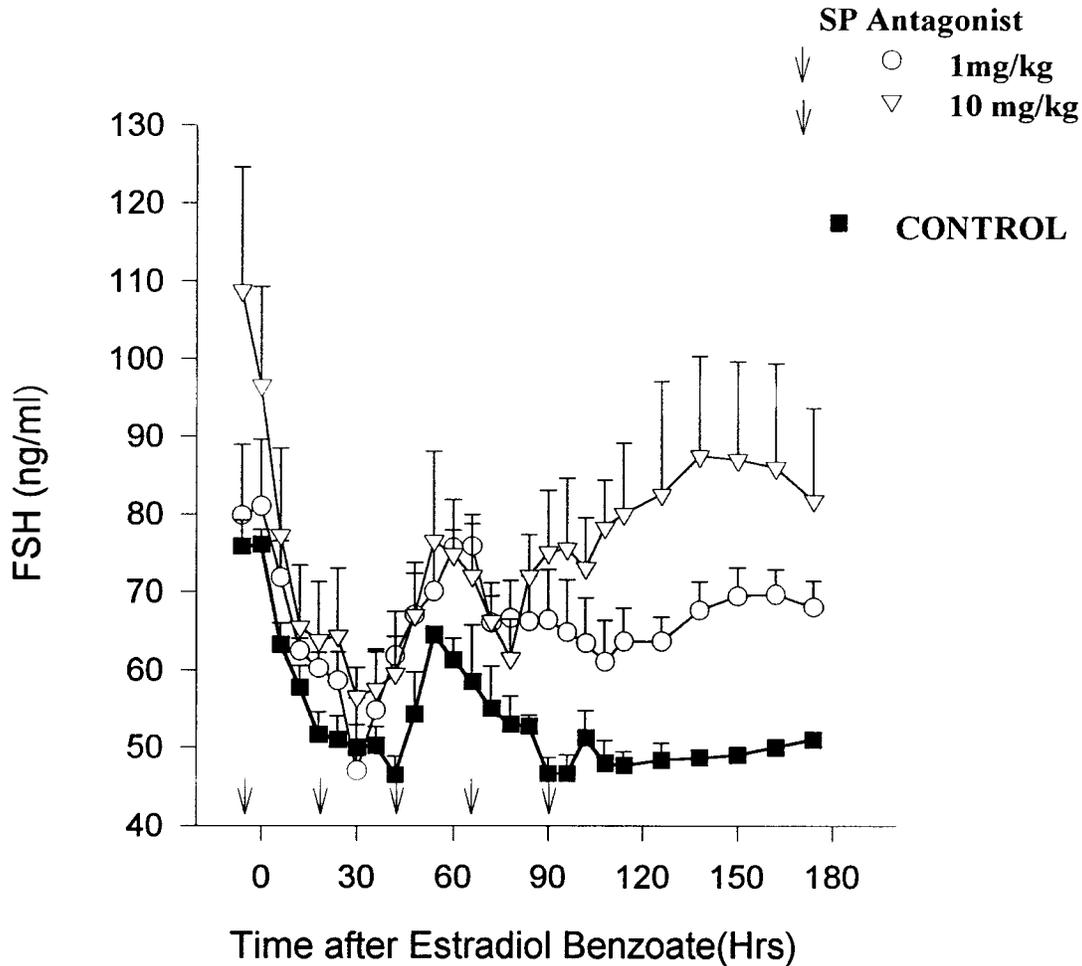


Fig. 5. Pattern of mean (\pm S.E.M.) plasma concentrations of FSH in blood samples taken up every 6 hr to the time of 174 hr, from OVX cynomolgus monkeys treated with one injection of E₂B (50 μ g/kg) at 0 hr and with five intragastric administrations of 1 mg/kg or 10 mg/kg of RPR 100893 or vehicle, at -6, 18, 42, 66, and 90 hr.

nucleus level in the monkey (Fardin et al., 1994b). In the present experiment, assuming a molecular weight of 561.73, a plasma concentration of 60 ng/ml represents a level of 107 nM and a concentration of 200 ng/ml represents a level of 356 nM. It shows that at least the plasma concentration of 60 ng/ml is in agreement with receptor occupation.

Another interesting finding of this study is the fact that the treatment of the OVX monkey with the human NK₁ receptor antagonist leads to a small but significant increase in plasma SP levels, showing a possible correlation between the occupancy of the NK₁ receptor and the secretion of SP. The fact that the RPR 100893 elevates circulating levels of substance P is difficult to explain. A phenomenon similar to the one occurring after blockade of the postsynaptic dopaminergic receptor (Lidsky and Banerjee, 1993; Seeman and Van Tol, 1994) which leads

to an augmentation of dopamine release could be envisioned. However, a similar phenomenon has not been described in the case of the SPergic transmission or for any peripheric transmission.

Elevated levels of SP were only observed for the smaller concentration of the NK₁ antagonist which is very similar to the IC₅₀ for the receptor. A down-regulation process could be initiated, with a return to control values for plasma SP, for the higher concentration of the NK₁ antagonist which gives rise to NK₁ plasma values close to 10 times the IC₅₀ (see above). However, the increase in circulating levels of SP observed after the administration of the smaller amount of the RPR 100893 could be a consequence of many inputs to the systemic system including a local release from peripherally situated nerve terminals spilling over to the bloodstream, or of an endocrine activity arising from the brain, the pituitary or

the gastrointestinal tract being tonically under the control of SP-mediated inhibition.

Previous studies (Wormald et al., 1989) suggest that it is very likely that the action of the human NK₁ receptor antagonist is exerted at the AP level. However, an action at the hypothalamic level cannot be completely excluded since administration of ¹⁴C-RPR 100893 radioactivity really crosses the blood barrier and is extensively plasma protein-bound (results not published). Also, it was shown that oral administration of RPR 100893 inhibits up to 50% of the binding of ³H-SP at the striatum level in the guinea pig (Flamand and Fardin, 1995). Furthermore, it was also shown that RPR 100893 administered by the i.v. route decreases c-fos expression in trigeminal nucleus caudalis following noxious chemical meningeal stimulation (intracisternal capsaicin injection) in the guinea pig (Cutrer et al., 1995), suggesting an action of the NK₁ antagonist at the brain level. In addition, the stereoisomer (RPR 103253) of the RPR 100893 did not reduce the number of positive cells in the trigeminal nucleus caudalis (Cutrer et al., 1995). Also, the stereoisomer does not block the neurogenic plasma protein extravasation within guinea pig dura mater and conjunctiva (Lee et al., 1994). These data show that the nonpeptide antagonist RPR 100893 does not display nonspecific side effects as does the activation of Ca⁺⁺ channels.

In summary, our data clearly show that NK₁ receptors are part of the hypothalamo-AP integrative network which is activated by 17β-estradiol for the LH and FSH release in the OVX monkey. It remains to be determined the extent of the stimulatory effect of the NK₁ antagonist administered during the physiological rise of 17β-estradiol which occurs during the follicular phase, on preovulatory LH and FSH surges, the subsequent ovulation and progesterone production during the luteal phase, in the normally menstruating monkey.

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