17. α-Alkylation of Amino Acids without Racemization. Preparation of Either (S)- or (R)-α-Methyldopa from (S)-Alanine

by Dieter Seebach^{*}, Johannes D. Aebi¹), Reto Naef²) and Theodor Weber¹)

Laboratorium für Organische Chemie der Eidgenössischen Technischen Hochschule, ETH-Zentrum, Universitätstrasse 16, CH-8092 Zürich

(29.VIII.84)

Enantiomerically pure *cis*- and *trans*-5-alkyl-1-benzoyl-2-(*tert*-butyl)-3-methylimidazolidin-4-ones (1, 2, 11, 15, 16) and *trans*-2-(*tert*-butyl)-3-methyl-5-phenylimidazolidin-4-one (20), readily available from (S)-alanine, (S)-valine, (S)-methionine, and (R)-phenylglycine are deprotonated to chiral enolates (*cf*. 3, 4, 12, 21). Diastereoselective alkylation of these enolates to 5,5-dialkyl- or 5-alkyl-5-arylimidazolidinones (5, 6, 9, 10, 13a-d, 17, 18, 22) and hydrolysis give α -alkyl- α -amino acids such as (R)- and (S)- α -methyldopa (7 and 8a, resp.), (S)- α methylvaline (14), and (R)- α -methyl-methionine (19). The configuration of the products is proved by chemical correlation and by NOE ¹H-NMR measurements (see 23, 24). In the overall process, a simple, enantiomerically pure α -amino acid can be α -alkylated with retention or with inversion of configuration through pivalaldehyde acetal derivatives. Since no chiral auxiliary is required, the process is coined 'self-reproduction of a center of chirality'. The method is compared with other α -alkylations of amino acids occurring without racemization. The importance of enantiomerically pure, α -branched α -amino acids as synthetic intermediates and for the preparation of biologically active compounds is discussed.

In [1], we have shown that simple amino acids such as (S)-alanine, (S)-valine, (R)-phenylglycine, (S)-phenylalanine, and (S)-methionine can be converted selectively to imidazolidinones such as 1 and 2 of either *cis*- or *trans*-configuration. We now show that these imidazolidinones are deprotonated to synthetically useful chiral enolates.

A) Reactions of the Chiral Imidazolidinone Enolates with Electrophiles. – As a first example, the preparation of (R)- or (S)- α -methyldopa (= 2-amino-2-methyl-3-(3,4-di-hydroxyphenyl)propionic acid) from (S)-alanine is described (Scheme 1). Solutions of the imidazolidinones (1 or 2) in THF were treated at dry-ice temperature with a slight excess of lithium diisopropylamide (LDA). Bright orange-red-colored solutions of the enantiomeric enolates (3, 4) were formed, which were combined with 3,4-dimethoxybenzyl bromide. Rapid decolorization indicated the progress of the alkylation step furnishing the enantiomeric 5,5-disubstituted imidazolidinones (5, 6, ca. 60%). We did not detect more than one diastereoisomer by HPLC of the crude products. Within experimental error, the two isomers 5 and 6 had identical physical properties such as melting points, IR, and NMR spectra, but an opposite sense of specific rotation.

The heterocyclic ring *and* the phenolic methyl-ether groups of the enantiomer **6** were cleaved by heating for 4 hours in 6N HCl at 180° in a sealed tube. Isolation of the α -methyldopa thus produced caused considerable problems due to the known [2a] sen-

¹) Part of the projected Ph. D. theses of J. D. A. and Th. W., ETH Zürich.

²) Part of the Dissertation No. 7442 of R.N., ETH Zürich, 1983.



sitivity of this substance towards oxygen (air!), especially under basic conditions. It appeared that the MeNH₂ formed in the ring cleavage as a by-product catalyzed decomposition of α -methyldopa. Therefore, the crude product **8a** was acetylated to the crystalline N,O,O-triacetyl derivative **8b**.

The enantiomer of 6 was also cleaved to give (R)- α -methyldopa $(5 \rightarrow 7)$. The specific rotations of 5 and 6, and the comparison of the specific rotation of 8b with that of authentic material [2b] are definitive proof for the diastereoisomeric and enantiomeric purities of the imidazolidinones 1 and 2. Thus, the overall conversion of (S)-alanine to

methyldopa has been achived with retention³) of configuration through the *cis*-isomer and with inversion³) through the *trans*-isomer. The enolates (3, 4) were alkylated from the face opposite to the *t*-Bu group (relative topicity lk [4])⁴).



³) For the definition of the use of the terms inversion and retention of configuration in cases like this, see [3a], especially footnote 21 therein (the paper mentioned has appeared in the meantime, *cf.* [3b]).

⁴) It is probably also possible to prepare 5 and 6, and thus α-methyldopa by methylation of dopa itself. The cis-imidazolidinone (1, C₆H₅CH₂ instead of CH₃) from (S)-phenylalanine could be prepared, albeit with ca. 50% racemization (enantiomerically pure (2R,5S)-1-benzoyl-5-benzyl-2-(*tert*-butyl)-3-methylimidazolidin-4-one has an [α]²⁵_D = -27°, m.p. 193°). A partially racemized sample was allylated (CH₂=CH--CH₂Br) in 81% yield to give diastereomerically pure u-1-benzoyl-5-benzyl-2-(*tert*-butyl)-3-methyl-5-(2-propen-1-yl)imidazolidin-4-one.

The (R)-enolate 3 derived from (S)-alanine was also ethylated (\rightarrow 9), and its enantiomer 4 benzylated (\rightarrow 10). Of the other imidazolinones reported in [1], we have alkylated⁴) those derived from (S)-valine (see 11), (S)-methionine (see 15, 16), and (R)-phenylglycine (see 20). In the case of the derivative 11, we first determined the stereochemical course of the deprotonation/deuteration by quenching the lithium enolate 12 with CH₃OD at -78° . This gave the deuterated product 13a with retention of configuration (ca. 70% yield, \geq 90% D-incorporation, \geq 95% (*R*,*S*)-diastereoisomer (by ¹H-NMR)). Thus, again, the attack of the electrophile D^+ has taken place cleanly from the (*Re*)-face of the enolate 12 double bond, opposite to the t-Bu group. Methylation, ethylation, and allylation of 12 gave single diastereomers 13b, 13c, and 13d, respectively, which are drawn with (5S)-configuration in the *Formulae*, and of which 13b was hydrolyzed to (S)- α -methylvaline (14) by heating its solution in 20% aq. HCl to 180° in a sealed tube for 4 hours. For configurational assignments, see Section B. The cis- and trans-substituted heterocycles 15 and 16 from (S)-methionine were deprotonated to the enantiomeric enolates and alkylated to give the geminally disubstituted derivatives 17 and 18, respectively; hydrolysis of 18 as described above furnished (R)- α -methylmethionine (19).

A special case is that of the non-acylated *trans*-substituted heterocycle 20, obtained from (*R*)-phenylglycine. Due to the additionally acidifying effect of the phenyl group of 20, and hence the increased stability of the enolate, it was possible to deprotonate this imidazolidinone without 'protection' of the NH-group and to ethylate the resulting enolate 21 in 80% yield and with high diastereoselectivity $(\rightarrow 22)^5$).

The imidazolidinone enolates described here are surprisingly stable. The orange to dark-red-colored solutions can be briefly warmed up to at least 0° without deterioration. The color of the solutions can be used as an indicator in reactions with electrophiles: it disappears or turns to a light yellow at -75° instantaneously when CD₃OD, CD₃COOD, or benzaldehyde are added, with allyl and benzyl bromides it takes a few minutes that the color fades away at the same temperature, while MeI and EtI require warming to temperatures between -50° and -30° . The yields of the alkylations range from 70 to 90%⁶). The imidazolidinone enolates appear to be more basic and less nucleophilic towards carbonyl derivatives than the corresponding enolates of dioxolanones [8] and oxazolidinones [3a] derived from α -hydroxy acids and from proline, respectively. Thus, the enolate **12** was quantitatively protonated upon addition of acetone, while it gave the adduct with the non-enolizable benzaldehyde⁷) in good yield.

⁵) The benzoylation of the other imidazolidinones 1, 2, 11, 15, and 16 does not only serve as a protection but should also acidify the neighboring CH-group: it is known that non-enolizable carboxamides with sterically protected but electronically effective carbonyl groups are readily deprotonated at the N-CH-position; for reviews, see [5-7].

⁶) Alkylating reagents such as secondary or long chain alkyl halides or sulfonates have not been tested in these reactions, as yet.

⁷) This and other adducts of imidazolidinone enolates and carbonyl derivatives will be described elsewhere. We expect the enolate **21** to be much less basic than the analogues with N-benzoyl groups, because a proline-derived enolate with nonacylated NH₂-group [3a] and dioxolanone enolates bearing atoms with 'free electron pairs' on the enolate β -C-atom [8] are also nonbasic, but highly nucleophilic.

B) Assignment of the Configuration of the Products 9, 10, and 22 by NOE ¹H-NMR-Measurements. – The relative and absolute configurations of the α -methyldopa derivatives 5 and 6, and of the valine derivative 13a were assigned by chemical correlation (see above). We have determined the relative configuration of three further 5,5-disubstituted imidazolidinones from nuclear *Overhauser* effects (NOE).

The NOE were preferably measured with a 360-MHz instrument in (D_6) benzene for better separation of the important ¹H-NMR signals in these experiments. Thus, upon irradiation of 9 with the resonance frequency of the *t*-Bu group, a positive NOE was observed at the signal of the CH₃-group in 5-position, indicating the *cis*-relationship of these two groups, see 23 (R¹ = CH₃). Likewise, in the ¹H-NMR spectrum of the phenylglycine-derived compound 22, the intensity of the signal of the *ortho*-protons in the phenyl group was enhanced by irradiating with the frequency of the *t*-Bu protons, see 23 (R¹ = C₆H₃).



Several of the ¹H-NMR spectra of 5,5-disubstituted imidazolidinones appeared very complex at first sight. In some cases, most signals were broad and unresolved, in others, two full sets of signals were present. Thus, a CDCl₃ solution of the alanine-derived compound **10**, which is nicely crystalline with a sharp m.p. at 112°, and which was a single, pure compound according to HPLC, was proved to contain two isomers by ¹H-NMR spectroscopy. Irradiation with the frequencies of the *t*-Bu protons caused a positive NOE in the signals from *both* CH₃-C(5), but only in those from the *ortho*-protons of phenyl of *one* of the isomers! This indicates that in both isomers, the *t*-Bu and the CH₃-groups are *cis* to each other, and that we are dealing with the (*E/Z*)-isomers **24a** and **24b** of the same *l*-diastereoisomer. Indeed, corresponding signals of the spectrum coalesce near the boiling point of (D₆)DMSO (*ca.* 180°).

In analogy with the established cases, the absolute and relative configurations of the other 5,5-disubstituted imidazolidinones (and of the products of their hydrolysis) are assumed without proof to be such that the substituent introduced in the reaction of the enolate with the electrophile is in a *trans*-position with respect to the *t*-Bu group, as drawn in the corresponding *Formulae* and as specified in the *Exper. Part.*

C) Discussion and Conclusions. – The present results show that α -amino acids can be α -alkylated without racemization, see *Scheme 2* for the transformation with inversion of configuration³); thus, a (S)- α -amino acid is, for instance, converted to the *trans*-imidazolidinone [1], a pivalaldehyde N,N-acetal with two asymmetric centers; this acetal is deprotonated to a nonracemic lithium enolate, which in turn reacts with electrophiles highly diastereoselectively, *i.e.* the acetal center exerts a strong asymmetric induction in the formation of the new center of chirality; finally, hydrolysis of the imidazolidinone with cleavage of the acetal and amide bonds completes the overall





alkylation of the amino acid. Since no chiral auxiliary is employed in the entire sequence, this was called a self-reproduction of the center of chirality. The only nonracemic compound used is the original α -amino acid. It is possible to perform this sequence of reactions also through the less readily available imidazolidinones of *cis*-configuration, *i.e.* with retention of configuration³); furthermore, many α -amino acids are now commercially available in both enantiomeric forms. The 'auxiliary' which is necessary for the enantioselective overall conversion is pivalaldehyde, a by-product of the industrial hydroformylation of 2-methylpropene (isobutylene). We have realized similar sequences, using the same auxiliary, for the α -alkylation of α -hydroxy- and α -mercaptocarboxylic acids (see **25** and [8]), of proline (see **26**, X = CH₂, and [3a]), and of cysteine and serine (see **26**, X = S, and **27**, and [9] [10], resp.).



⁸) The configuration of the enolate double bonds shown in the *Formulae* **27**, **29**, and **30** has not been established; the drawing is arbitrary, as long as structural information is missing; see [14].

Previous methods of preparing enantiomerically pure α -branched α -amino acids had to rely on chiral auxiliaries such as other α -amino acids in *Schöllkopf*'s procedure (see **28** and [11] [12]) or 2-hydroxypinan-3-one as employed by *Viallefont et al.* (see **29** and [13]). For practical purposes and large scale preparations, these auxiliaries would have to be recycled⁹).

We have previously shown that chiral dihydroxy-carboxylic acids such as tartaric acid [15], as well as β -hydroxy α -amino acids such as threonine, allothreonine, and phenylserine (see 30, $R = CH_3$ and C_6H_5 , resp., and [16]) can also be alkylated stereoselectively. Work on the α -alkylation of other functionalized amino acids (tyrosine, tryptophane, aspartic acid, glutamic acid, lysine, ornithine, arginine, histidine etc.) is in progress in our laboratory. Thus, an increasing number of enantiomerically pure compounds (EPC) with persubstituted C-atoms become available as starting materials for EPC-syntheses of natural products and of physiologically active compounds. This is important not only for the synthetic chemist: more and more α -branched α -amino acids are now being discovered as minor components of peptides or are isolated from other natural sources¹⁰). They have been shown to be biologically active such as (S)- α methyldopa (8) which exhibits hypotensive activity and which inhibits the decarboxylation of (S)-dopa by mammalian decarboxylase, while the (R)-enantiomer is totally inactive [18]. Some hydantoins derived from α -branched amino acids are antiepileptica [19]. On the other hand, the highly toxic α -aminocyclopentanecarboxylic acid has been shown to have carcinostatic properties¹¹) while derivatives of α -methylmethionine and ethylalanine trigger tumor growth [21]. Finally, it is to be expected that incorporation of branched amino acids at certain positions of physiologically active oligopeptides will be possible with preservation of activity and simultaneous protection against cleavage by peptidases. Our method provides ready access to a large variety of α -branched amino acids for tests and further elaboration and application.

We thank the same persons, firms, and institutions as stated in [1] for generous help and support during these investigations.

Experimental Part

General. See [1]. Melting points (m. p.) and boiling points (b. p.) are uncorrected. Reagents and solvents were purified in the usual way. THF was distilled under Ar over K. All reactions involving Li derivatives were carried out under anh. conditions in an Ar atmosphere [22]. The diastereoisomeric composition of crude products was determined by ¹H-NMR. The anisochronous ¹H-NMR signals of the minor diastereoisomer are given in *italics*. Flash chromatography was performed according to the method described by *Still et al.* [23].

A) General Procedure for Reactions of the Enolates 25 with Various Electrophiles. – Unless noted otherwise, 1.1 equiv. of a 1 M LDA solution in THF/hexane 1:3 was added to a 0.17m solution of the imidazolidinone derivative in THF at -78° (\rightarrow deep red). After 30 min, 1.2 equiv. of the electrophile were added, and the temp.

⁹) This is an especially problematic task in the case of the highly selective reaction used by *Schöllkopf et al.* [11] [12], because two amino-acid esters, the auxiliary one and the branched one, have to be separated from each other; this is done by fractional distillation or by preparative gas chromatography following the final step!

¹⁰) See the recent reviews [17].

¹¹) Leucinostatin A, a novel nonapeptide with antitumor activity, contains three 2-amino-2-methylpropionicacid units [20].

was allowed to warm to 0° within 2 h. The resulting light yellow solution was poured into half-sat. NH_4Cl soln. and extracted twice with Et_2O (200 ml). The org. phase was washed once with 150 ml of dist. H_2O and dried (MgSO₄). The solvent was then removed *in vacuo*.

B) General Procedure for Hydrolysis of the 5,5-Dialkylated Imidazolidinones. – The purified 5,5-dialkylated imidazolidinone (1 g) was mixed with 20 ml of aq. 6N HCl. The sealed tube was heated at 175–185° for 4 h in a steel tube after which the heterogeneous mixture was washed with 15 ml of CH_2Cl_2 . Concentration under reduced pressure gave the crude amino-acid hydrochlorid.

C) General Procedure for Production of the Free Amino Acid. – The salt was dissolved in 10 ml of H_2O and adsorbed on 30 g of acidic *Dowex 50 WX8*. The resin was washed with dist. H_2O until neutral, and then the free amino acid was eluted with 1.3N aq. NH₃. The solvent was removed *in vacuo*. Complete removal of NH₃ was accomplished by redissolving the substance in H_2O and concentrating in a rotatory evaporator. Finally, drying for 24 h under high vacuum (50°) provided pure amino acids.

(2R,5R)-1-Benzoyl-2-(tert-butyl)-5-(3',4'-dimethoxybenzyl)-3,5-dimethylimidazolidin-4-one (5). Following Procedure A, 2.74 g (10 mmol) of 1 and 2.8 g (12 mmol) of 3,4-dimethoxybenzyl bromide dissolved in 15 ml of THF provided, after flash chromatography (Et₂O/pentane 5:1), 2,47 g (57%) of 5 as white powder. The following modification improved the yield by 15%: Using a *Teflon* tube as a cannula, the enolate solution (0.1M, -78°) was transferred dropwise to the 0.31M 3,4-dimethoxybenzyl bromide solution (-78°). M.p. 165°, $[\alpha]_{15}^{25} = -78.0$ (c = 0.5, CHCl₃). IR (KBr): 2960m, 1710s, 1640s, 1520m, 1370m, 1260m, 1240m, 1135m. ¹H-NMR (CDCl₃, mixture of 2 rotamers): 7.80–6.67 (m, 8 arom, H); 5.26/4.92 (s, H–C(2)); 3.90/3.51 (s, CH₃O); 3.80–3.00 (m, C₆H₅CH₂); 2.97/2.60 (s, CH₃–C(5)); 1.00/0.77 (s, (CH₃)₃C). MS: 424 (1, M^{+}), 367 (12), 151 (18), 105 (100), 77 (22). For the microanalysis, a sample was crystallized from MeOH/Et₂O at -78° . ¹H-NMR; presence of 0.5 equiv. of MeOH in the crystal. Anal. calc. for C_{25.5}H₃₄N₂O_{4.5} (440.53): C 69.52, H 7.77, N 6.36; found: C 69.68, H 7.47, N 6.46.

(2S,5S)-1-Benzoyl-2-(tert-butyl)-5-(3',4'-dimethoxybenzyl)-3,5-dimethylimidazolidin-4-one (6). Following Procedure A, 2.74 g (10 mmol) of 2 and 2.8 g (12 mmol) of 3,4-dimethoxybenzyl bromid dissolved in 15 ml of THF provided after flash chromatography (Et₂O/pentane 5:1) 2.64 g (61%) of 6 as white powder. $[\alpha]_D^{25} = +74.9$ (c = 0.4, CHCl₃). Other physical data are identical with those of 5.

(2S)-N-Acetyl-3-(3',4'-diacetoxyphenyl)-2-methylalanine (**8b**). Following Procedure B, 4.69 g (11.05 mmol) of **6** in 30 ml of aq. 6N HCl provided 3.67 g of a mixture of ammonium salts. To remove the MeNH₂ this crude product was dissolved in 20 ml of pyridine and evaporated to dryness *in vacuo* (0.01 Torr). For acetylation of **8a**, the residue was dissolved in 60 ml of Ac₂O/pyridine 2:1 and heated at 95° for 3 h¹²). The dark solution was concentrated *in vacuo*. The residual oil was dissolved in 5 ml of acetone, 10 ml of H₂O and 1 ml of 2.5M HCl¹³). After evaporation, the oil was dissolved in 20 ml of EtOH and poured into 200 ml of H₂O/Et₂O 1:3. The H₂O phase was extracted twice with Et₂O (100 ml). Then, the org. phase was evaporated and dried *in vacuo*. Crystallization from acetone/pentane gave 2.44 g (56%) of **8b**. $[\alpha]_D^{25} = -74.7$ (c = 0.73, MeOH))¹⁴). One further crystallization gave 1.23 g (33%) of pure **8b**. M. p.: 178.0–179.0°. $[\alpha]_D^{25} = -93.9$ (c = 1.04, MeOH)¹⁴) ([2b]: m.p. 180.0–181.0°; $[\alpha]_D^{25} = -94.2$ (c = 0.716, MeOH)). ¹H-NMR (CD₃OD): 7.20–6.91 (*m*, 3 arom. H); 4.85 (br. *s*, NH, COOH); 3.46, 3.11 (*AB*, J = 13.5, 2H–C(3)); 2.21 (s, 2 OCOCH₃); 1.90 (s, NHCOCH₃); 1.40 (s, CH₃–C(2)).

(2R)-3-(3',4'-Dihydroxyphenyl)-2-methylalanine (= (R)- α -Methyldopa; 7). Following Procedure B, 0.6 g (1.41 mmol) of 5 in 20 ml of aq. 6N HCl provided 0.47 g of 7 as the methylammonium salt. ¹H-NMR (D₂O; HDO = 4.8 ppm): 6.91-6.50 (m, 3 arom. H); 3.10, 2.86 (AB, J = 13.5, 2H-C(3)); 2.53 (s, CH₃N); 1.55 (s, CH₃-C(2)).

(2R,5R)-1-Benzoyl-2-(tert-butyl)-5-ethyl-3,5-dimethylimidazolidin-4-one (9). Following Procedure A, 2.74 g (10 mmol) of 1 and 1 ml (12 mmol) of EtI provided, after flash chromatography (Et₂O/pentane 2:1), 2.72 g (90%) of 9 as colorless crystals. For analytical purposes 9 crystallized from Et₂O/pentane (ds >95%). M.p. 122.5-123.5°. $[\alpha]_{25}^{D5} = +16.5$ (c = 1.43, CHCl₃). IR (CHCl₃): 2970m, 1680s, 1630s, 1355s, 1090w. ¹H-NMR (CDCl₃): 7.66-7.36 (m, 5 arom. H); 5.74 (s, H-C(2)); 3.10 (s, CH₃N); 1.67-1.56 (q, J = 9, CH₃CH₂); 1.53 (s,

¹²) All manipulations were carried out under an Ar [2].

¹³) The acid was added to accelarate hydrolysis of the azalactone.

¹⁴) The ¹H-NMR spectra shows that 1 equiv. of acetone is present in the crystal. For anal. measurements, the crystals were dissolved several times in CH₂Cl₂ and the solvent evaporated. Finally, **8b** was precipitated with pentane and dried *in vacuo*.

CH₃-C(5)); 1.10 (s, (CH₃)₃C); 0.60 (t, J = 9, CH₃CH₂-C(5)). MS: 287 (1, M^+ - 15), 245 (25), 105 (100), 77 (27), 42 (10). Anal. calc. for C₁₈H₂₆N₂O₂ (302.34): C 71.49, H 8.67, N 9.26; found: C 71.38, H 8.57, N 9.38.

(2S,5S)-*1-Benzoyl-5-benzyl-2-(* tert-*butyl)-3,5-dimethylimidazolidin-4-one* (10). Following *Procedure A*, 2.74 g (10 mmol) of **2** and 1.5 ml (12 mmol) of benzyl bromide provided, after flash chromatography (Et₂O/pentane 5:1), 2.65 g (73%) of 10 as colorless crystals (ds > 95%). M.p.: 112°. $[\alpha]_{D}^{25} = +67.7$ (c = 1.0, CHCl₃). IR (KBr): 2990m, 1700s, 1642s, 1400m, 1375s. ¹H-NMR (C₆D₆; mixture of two rotamers): 7.50–6.90 (m, 10 arom. H); 5.17/4.66 (s, H–C(2)); 3.98, 3.34/2.61, 2.53 (*AB*, J = 14, C₆H₅CH₂); 2.49/2.28 (s, CH₃N); 1.91/1.44 (s, CH₃-C(5)); 0.76/0.42 (s, (CH₃)₃C). MS: 349 (1, $M^+ - 15$), 307 (100), 306 (33), 305 (94), 77 (90), 57 (22). Anal. calc. for C₂₃H₂₈N₂O₂ (364.45): C 75.79, H 7.74, N 7.68; found: C 75.71, H 7.86, N 7.57.

(2R,5S)-1-Benzoyl-2-(tert-butyl)-5-isopropyl-3-methyl- $(5^{-2}H)$ imidazolidin-4-one (13a). Following Procedure A, 2.74 g (10 mmol) of 11 and 1.7 ml (30 mmol) of CH₃COOD provided a max. deuteration of 20%. At -30° , the enolate-solution of 12 was evaporated under high vacuum, and the resinous residue was washed several times with hexane yielding 12 as powdery, orange solid. A sample was quenched in the NMR tube at 0° with CD₃OD. The degree of deuteration was $\geq 95^{\circ}$ (79% ds). The molar ratio of imidazolidinone 13a to THF was exactly 1:1. The main part of 12 was suspended at -78° in hexane and quenched with CH₃OD. Workup as described in Procedure A gave 2.05 g (68%) of 13a with a degree of deuteration of $\geq 90^{\circ}$ (ds > 95%). M.p. 116°. $[\alpha]_D^{25} = +24.8$ (c = 1.2, CHCl₃). ([1]: m.p. 112°; $[\alpha]_D^{25} = +22.4$ (c = 1.7, CHCl₃)). IR (CHCl₃): 2965m, 1685s, 1635s, 1362s, 1295w. ¹H-NMR (CCl₄): 7.46-7.13 (m, 5 arom. H); 5.33 (s, H-C(2)); 3.86 (< 10%, d, J = 10, H-C(5)); 2.97 (s, CH₃N); 2.06-1.46 (m, (CH₃)₂CH); 1.03 (s, (CH₃)₃C); 1.00 (d, J = 6, CH₃CHCH₃). Anal. calc. for C₁₈H₂₅DN₂O₂ (303.38): C 71.26, H 8.96, N 9.32; found: C 71.47, H 8.70, N 9.23.

(2R,5S)-1-Benzoyl-2-(tert-butyl)-5-isopropyl-3,5-dimethylimidazolidin-4-one (13b). Following Procedure A, 3.02 g (10 mmol) of 11 and 0.75 ml (12 mmol) of MeI provided, after flash chromatography (Et₂O/pentane 5:1), 2.18 g (70%) of 13b as colorless crystals (ds > 95%). M.p. 95.0-97.5°. $[\alpha]_{25}^{D5} = +38.7$ (c = 1.38, CHCl₃). IR (CHCl₃): 2955m, 2895w, 1695s, 1650s, 1410s, 1271m, 1122m, 1102m, 1036w, 935w, 708w. ¹H-NMR (CDCl₃): 7.43 (s, 5 arom. H); 5.46 (s, H-C(2)); 3.03 (s, CH₃N); 2.26-1.76 (m, (CH₃)₂CH); 1.37 (s, CH₃C-C(5)); 1.15 (d, J = 7, CH₃CHCH₃); 1.12 (d, J = 7, CH₃CHCH₃); 1.02 (s, (CH₃)₃C). MS: 301 (0.3, $M^+ - 15$), 260 (6), 259 (32), 106 (8), 105 (100), 77 (14), 42 (5). Anal. calc. for C₁₉H₂₈N₂O₂ (316.45): C 72.11, H 8.92, N 8.85; found: C 72.25, H 8.76, N 8.84.

(2R,5S)-*1-Benzoyl-2-(*tert-*butyl)-5-ethyl-5-isopropyl-3-methylimidazolidin-4-one* (13c). Following *Procedure A*, 3.02 g (10 mmol) of 11 and 1 ml (12 mmol) of EtI provided, after one crystallization (Et₂O/pentane), 2.67 g (81%) of diastereoisomerically pure 13c as white crystals. M.p. 112°. [α]₂₅²⁵ = +39.9 (c = 1.3, CHCl₃). IR (CHCl₃): 2960s, 1675s, 1630s, 1450m, 1350s, 1090m. ¹H-NMR (CDCl₃; mixture of two rotamers): 7.43 (s, 5 arom. H); 5.56/5.43 (s, H–C(2)); 3.03 (s, CH₃N); 2.20–1.70 (m, (CH₃)₂CH); 1.33–0.56 (m, (CH₃)₂CH, (CH₃)₃C, CH₃CH₂CH, (CH₃)₃C, CH₃CH₂). MS: 331 (46, M ⁺ + 1), 274 (74), 273 (95), 105 (90), 77 (100), 57 (15). Anal. calc. for C₂₀H₃₀N₂O₂ (330.44): C 72.69, H 9.15, N 8.48; found: C 72.57, H 9.02, N 8.39.

(2R,3S)-5-Allyl-1-benzoyl-2-(tert-butyl)-5-isopropyl-3-methylimidazolidin-4-one (13d). Following Procedure A, 3.02 g (10 mmol) of 11 and 1 ml (12 mmol) of allyl bromide provided, after a bulb-to-bulb distillation, 3.05 g (89%) of diastereoisomerically pure 13d as a colorless resin. B.p. 190°/0.1 Torr. $[\alpha]_{25}^{25} = -5.94$ (c = 0.41, CHCl₃). IR (CHCl₃): 2970m, 1690s, 1635s, 1365m, 1090w. ¹H-NMR (CCl₄): 7.34 (s, 5 arom. H); 5.73–4.86 (m, CH₂=CHCH₂, H–C(2)); 2.83 (s, CH₃N); 2.80–2.43 (m, (CH₃)₂CH); 2.33–1.56 (m, CH₂=CHCH₂); 1.40–0.66 (m, (CH₃)₂CH, (CH₃)₃C). MS: 341 (2, $M^{+} - 1$), 285 (15), 105 (100), 77 (25). Anal. calc. for C₂₁H₃₀N₂O₂ (342.45): C 73.65, H 8.83, N 8.18; found: C 73.25, H 8.70, N 8.25.

(2S)-2-Amino-2,3-dimethylbutanoic Acid (= (S)- α -methylvaline; 14). Following Procedure B, 1.36 g (4.3 mmol) of 13b in 30 ml of aq. 6N HCl provided the crude hydrochloride of 14. Cation-exchange chromatography by Procedure C gave 0.564 g (95%) of 14 as colorless crystals. Dec. 215°. [α]_D²⁵ = -4.6 (c = 1.15, 0.2N HCl). IR (KBr): 3440m (br.), 3300-2200s (br.), 1610s, 1530m, 1465m, 1405s, 1370s, 1335s, 1320s, 1251w, 1230w, 1175w, 1160m, 1078w, 902m, 861m, 790w, 768m, 631w. ¹H-NMR (D₂O, HDO = 4.8 ppm): 2.19 (sept., J = 6.3, H-C(3)); 1.50 (s, CH₃-C(2)); 1.04 (d, J = 6.3, CH₃-C(3)); 1.00 (d, J = 6.3, 3H-C(4)). MS: 132 (1, M⁺ + 1), 131 (0.5, M⁺), 89 (7), 88 (100), 87 (5), 86 (74), 72 (6), 71 (11), 70 (14), 69 (19), 43 (18), 42 (59), 41 (19). Anal. calc. for C₆H₁₃NO₂ (131.18): C 54.94, H 9.99, N 10.68; found: C 54.85, H 10.02, N 10.57.

(2R,5S)-1-Benzoyl-2-(tert-butyl)-5-ethyl-3-methyl-5-(3'-thiabutyl)imidazolidin-4-one (17). Following Procedure A, 3.34 g (10 mmol) of 15 and 1 ml (12 mmol) of EtI provided, after flash chromatography (Et₂O/pentane 5:1), 3.04 g (84%) of 17 as colorless crystals (ds > 95%). M.p. 86°. $[\alpha]_{D}^{25} = +81.3$ (c = 1.0, CHCl₃). IR (KBr): 2975m, 1695s, 1640s, 1350m, 1330m, 1265m. ¹H-NMR (CDCl₃): 7.50 (br. s, 5 arom. H); 5.70 (s, H-C(2)); 3.06 (s, CH₃N); 3.00-1.50 (m, 3 CH₂); 2.03 (s, CH₃S); 1.10 (s, (CH₃)₃C); 0.66 (t, J = 6,

CH₃-C-C(5)). ¹³C-NMR (CDCl₃): 174.26 (*s*); 170.95 (*s*); 137.37 (*s*); 130.34 (*d*); 127.63 (*d*); 126.71 (*d*); 79.38 (*d*); 67.85 (*s*); 40.05 (*t*); 37.70 (*s*); 30.66 (*q*); 29.82 (*t*); 28.68 (*t*); 26.51 (*q*); 14.64 (*q*); 6.94 (*q*). MS: 362 (1, M^+), 306 (71), 305 (92), 111 (34), 106 (100), 105 (91), 77 (89), 42 (59), 41 (39). Anal. calc. for C₂₀H₃₀N₂O₂S (362.44): C 66.26, H 8.34, N 7.73; found: C 66.07, H 8.25, N 7.70.

(2S,5R)-1-Benzoyl-2-(tert-butyl)-3,5-dimethyl-5-(3'-thiabutyl)imidazolidin-4-one (18). Following Procedure A, 3.34 g (10 mmol) of 16 and 0.9 ml (15 mmol) of MeI provided, after one crystallization (CHCl₂/pentane), 2.36 g (66%) of 18 as slightly yellow crystals (ds > 95%). M.p. 99-107°. $[\alpha]_{25}^{25} = -76.1$ (c = 1.0, CHCl₃). IR (CHCl₃): 2970s, 1690s, 1640s, 1400m, 1300m, 1260m, 1115m, 1075m, 870m, 650m. ¹H-NMR (CDCl₃): 7.50 (s, 5 arom. H); 5.83 (s, H-C(2)); 3.84 (s, CH₃N); 3.00-2.00 (m, CH₂CH₂); 2.03 (s, CH₃S); 1.10 (s, CH₃-C(5), (CH₃)₃C). MS: 291 (100), 112 (10), 106 (29), 105 (86), 77 (64), 42 (15), 41 (11). Anal. cale. for C₁₉H₂₈NO₂S (348.51): C 65.48, H 8.09, N 8.03; found: C 65.35, H 8.04, N 8.12.

(2R)-2-Amino-2-methyl-5-thiahexanoic Acid (= (R)- α -Methylmethionine; **19**). Following the Procedures B and C, 1 g (2.8 mmol) of **18** in 25 ml of aq. 6N HCl provided 0.324 g (72%) of **19** as colorless crystals. M.p. 265–270°. [α] $_{D5}^{25} = -17.9$ (c = 0.7, 0.2N HCl). IR (KBr): 3450 (br.), 3300–2000s (br.), 1600s, 1455m, 1435m, 1400s, 1370m, 1305m, 1250m, 1120w, 890w, 550m. ¹H-NMR (D₂O, HDO = 4.80 ppm): 2.73–2.46 (m, 2H–C(4)); 2.22–1.93 (m, 2H–C(3)); 2.13 (s, CH₃S); 1.53 (s, CH₃–C(2)). MS: 163 (63, M^+), 146 (16), 118 (41), 89 (20), 88 (29), 75 (21), 71 (18), 70 (36), 61 (42), 57 (26), 46 (28), 45 (64), 43 (27), 42 (28), 41 (17), 31 (100), 27 (22), 18 (21). Anal. calc. for C₆H₁₃NO₂S (163.24): C 44.14, H 8.02, N 8.58; found: C 43.98, H 7.97, N 8.77.

(2S,5S)-2-(tert-Butyl)-5-ethyl-3-methyl-5-phenylimidazolidin-4-one (22). Following Procedure A, 2.23 g (10 mmol) of 20 and 1 ml (12 mmol) of EtI provided, after one crystallization at -20° (Et₂O/pentane), 2.08 g (80%) of 22 as slightly yellow crystals (ds > 95%). Addition of tris(*d*,*d*-dicampholylmethanato)europium(III) did not split signals in ¹H-NMR. M.p. 129°. [α]₂₅²⁵ = -6.76 (*c* = 2.2, CHCl₃). IR (CHCl₃): 3450*m* (br.), 2960*s*, 1670*s*, 1390*m*, 1075*m*. ¹H-NMR (CDCl₃): 7.81-7.20 (*m*, 5 arom. H); 4.25 (*s*, H–C(2)); 2.99 (*s*, CH₃N); 2.11-1.30 (*m*, CH₂); 2.05 (br. *s*, NH); 0.93 (*s*, (CH₃)₃C); 0.85 (*t*, *J* = 6, CH₃–C–C(5)). MS: 259 (1, *M*⁺ – 1), 203 (100), 134 (44), 104 (10), 42 (17). Anal. calc. for C₁₆H₂₄N₂O (260.35): C 73.81, H 9.29, N 10.76; found: C 73.86, H 9.37, N 10.83.

REFERENCES

- [1] R. Naef, D. Seebach, Helv. Chim. Acta 1985, 68, 135.
- [2] a) H. L. Slates, D. Taub, C. H. Kuo, N. L. Wendler, J. Org. Chem. 1964, 29, 1424; b) S. Yamada, S. Terashima, K. Achiwa, Chem. Pharm. Bull. Jpn. 1965, 13, 227; S. Terashima, K. Achiwa, S. Yamada, Chem. Pharm. Bull. Jpn. 1966, 14, 579.
- [3] a) D. Seebach, M. Boes, R. Naef, W.B. Schweizer, J. Am. Chem. Soc. 1983, 105, 5390; b) C.E. Wintner, J. Chem. Educ. 1983, 60, 550.
- [4] D. Seebach, V. Prelog, Angew. Chem. 1982, 94, 696; ibid. Int. Ed. 1982, 21, 654.
- [5] Review: D. Seebach, D. Enders, Angew. Chem. 1975, 87, 1; ibid. Int. Ed. 1975, 14, 15.
- [6] Reviews: P. Beak, D. B. Reitz, Chem. Rev. 1978, 78, 275; P. Beak, W.J. Zajdel, D. B. Reitz, ibid. 1984, 84, 471; N.G. Rondan, K.N. Houk, P. Beak, W.J. Zaidel, J. Chandrasekhar, P. v. R. Schleyer, J. Org. Chem. 1981, 46, 4108; D. B. Reitz, P. Beak, A. Tse, J. Org. Chem. 1981, 46, 4316, and ref. cited therein.
- [7] J.J. Lohmann, D. Seebach, M.A. Syfrig, M. Yoshifuji, Angew. Chem. 1981, 93, 125; ibid. Int. Ed. 1981, 20, 128. Review: D. Seebach, J.-J. Lohmann, M.A. Syfrig, M. Yoshifuji, Tetrahedron 1983, 39, 1863.
- [8] Review: D. Seebach, R. Naef, G. Calderari, Tetrahedron 1984, 40, 1313.
- [9] D. Seebach, Th. Weber, Tetrahedron Lett. 1983, 24, 3315; Helv. Chim. Acta 1984, 67, 1650
- [10] D. Seebach, J.D. Aebi, Tetrahedron Lett. 1984, 25, 2545.
- [11] Review: U. Schöllkopf, Tetrahedron 1983, 39, 2085; U. Schöllkopf, J. Nozulak, U. Groth, Tetrahedron 1984, 40, 1409.
- [12] Review: U. Schöllkopf, Topics Curr. Chem. 1983, 109, 65.
- [13] J. A. Bajgrowicz, B. Cossec, Ch. Pigière, R. Jacquier, Ph. Viallefont, Tetrahedron Lett. 1983, 24, 3721; cf. also: T. Oguri, T. Shioiri, S. Yamada, Chem. Pharm. Bull. Jpn. 1977, 25, 2287, and ref. cited therein.
- [14] D. Seebach, in 'The Robert A. Welch Foundation Conferences on Chemical Research. XXVII. Stereospecifity in Chemistry and Biochemistry'. Houston, Texas, Nov. 7–9, 1983, published in the proceedings of the above conference, *Welch* Foundation, Houston, 1984.
- [15] R. Naef, D. Seebach, Angew. Chem. 1981, 93, 1113; ibid. Int. Ed. 1981, 20, 1040.

- [16] D. Seebach, J.D. Aebi, Tetrahedron Lett. 1983, 24, 3311.
- [17] I. Wagner, H. Musso, Angew. Chem. 1983, 95, 827; ibid. Int. Ed. 1983, 22, 816; B. Witkop, Naturwissensch. Rundschau 1983, 36, 261.
- [18] A. Sjoerdsma, S. Udenfriend, Biochem. Pharmacol. 1961, 8, 164; S. M. Hess, R. H. Connamacher, M. Ozaki, S. Udenfriend, J. Pharmacol. Exp. Ther. 1961, 134, 129; C. C. Porter, J. A. Totaro, C. M. Leiby, *ibid.* 1961, 134, 139.
- [19] L.H. Goodson, I.L. Honigberg, J.J. Lehman, W.H. Burton, J. Org. Chem. 1960, 25, 1920; J.H. Poupaert, J. Adline, M.H. Claesen, P. De Laey, P.A. Dumont, J. Med. Chem. 1979, 22, 1140; A. Küpfer, P. Desmond, S. Schenker, R.A. Branch, Clin. Res. 1980, 28, 239 A; A. Küpfer, P. V. Desmond, S. Schenker, R.A. Branch, J. Pharmacol. Exp. Ther. 1982, 221, 590.
- [20] T.A. Connors, L.A. Elson, W.C.J. Ross, Biochem. Pharmacol. 1958, 1, 239; T.A. Connors, L.A. Elson, A. Haddow, W.C.J. Ross, Biochem. Pharmacol. 1960, 5, 108; Y. Mori, M. Tsuboi, M. Suzuki, K. Fukushima, T. Arai, J. Chem. Soc., Chem. Commun. 1982, 94.
- [21] J.A. Stock, in 'Ciba Foundation Symp. on Amino Acids with Antimetabolic Activity', Eds. G.E.W. Wolstenholme and C.M. O'Connor, J.&A. Churchill Ltd., London, 1958, p.89.
- [22] D. Seebach, T. Weller, G. Protschuk, A.K. Beck, M.S. Hoekstra, Helv. Chim. Acta 1981, 64, 716.
- [23] W.C. Still, M. Kahn, A. Mitra, J. Org. Chem. 1978, 43, 2923.