

Formation of Mineralized Bone Nodules by Rat Calvarial Osteoblasts Decreases With Donor Age Due to a Reduction in Signaling Through EP₁ Subtype of Prostaglandin E₂ Receptor

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Abstract The effects of prostaglandin E₂ (PGE₂) on the parameters for proliferation and differentiation were studied in calvarial osteoblast-like cells isolated from rats of various ages. In cells not treated with PGE₂, it was found that mineralized bone nodule (BN) formation, alkaline phosphatase (ALP) activity, and the incorporation rate of [³H]thymidine into the cells sharply decreased with the age of the cell donor at 6–50 weeks and then remained at a relatively constant level up to 120 weeks. Before studying the effects of PGE₂ on these parameters, we determined the change in the levels of PGE₂ produced by the untreated cells during the culture period and found that the endogenous PGE₂ reached a maximum on the 4th day of the culture, regardless of the cell donor age, followed by a sharp decrease. The endogenous production was blocked by pretreatment with a cyclooxygenase-2 (COX-2) inhibitor, NS-398, indicating the generation of PGE₂ through a COX-2 pathway. The area of BN was effectively suppressed by NS-398 in the cells from 10- to 35-week-old rats, whereas it was enhanced in the cells from 90- to 120-week-old rats. Treatment with PGE₂ markedly increased the BN formation and the ALP activity in the cells from 4- to 35-week-old rats (defined as young rats). By contrast, PGE₂ decreased [³H]thymidine incorporation into the cells from young rats. The area of BN and the ALP activity decreased significantly, whereas [³H]thymidine incorporation into the cells increased by 60–80% in the cells of 80- to 120-week-old rats (defined as aged rats). The stimulatory effects on the cell differentiation and the inhibitory effect on the proliferation in the cells from young rats was mimicked by an EP₁ agonist, 17-phenyl- ω -trinor PGE₂, while an EP₂/EP₄ agonist, 11-deoxy-PGE₁ and an adenylate cyclase activator, forskolin suppressed the differentiation and enhanced the proliferation regardless of the cell donor age. PGE₂, 11-deoxy-PGE₁ and forskolin, but not 17-phenyl- ω -trinor PGE₂ increased cyclic adenosine monophosphate (cAMP) production. Generation of inositol 1,4,5-triphosphate (IP₃) was stimulated by 17-phenyl- ω -trinor PGE₂ or PGE₂, but not by 11-deoxy-PGE₁ or forskolin increased cAMP production in the cells from young rats. By contrast, PGE₂ had little effect on IP₃ generation in aged rats. From the overall results, we concluded that PGE₂ exerts stimulatory and inhibitory effects on differentiation through the EP₁-IP₃ pathway and EP₂/EP₄-cAMP pathway, respectively, in the cells from young rats. The EP₁-IP₃ pathway seems to be inactive in the cells from aged rats. *J. Cell. Biochem.* 75:215–225, 1999. © 1999 Wiley-Liss, Inc.

Key words: osteoblasts; aging; cAMP; phosphoinositide turnover; prostaglandin E₂ (PGE₂); mineralized bone nodule

Prostaglandin E₂ (PGE₂) is a potent modulator of bone metabolism that modifies functions of osteoblasts as well as osteoclasts [Nijweide et al., 1986; Norrdin et al., 1990]. In vivo and in vitro studies showed that PGE₂ is largely involved in the regulation of bone formation [Chyun and Raisz, 1984; Yang et al., 1993], although its effect is variable, depending on the

experimental conditions employed [Suponitzky and Weinreb, 1998]. In vivo administration of PGE₂ to rats enhances cortical bone mass and activates intracortical bone remodeling [Jee et al., 1985, 1990]. The addition of PGE₂ to organ cultures of fetal rat calvaria causes stimulation of DNA synthesis in the periosteum and suppression of collagen synthesis in the central bone [Raisz and Koolemans-Beynen, 1974]. PGE₂-induced cellular responses in cultured osteoblastic cells have been studied most extensively in a osteoblastic cell line, MC3T3-E1, in which the effect of PGE₂ is differentiative at low

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concentrations and proliferative at high concentrations [Hakeda et al., 1986]. The biphasic effect of PGE₂ is attributed to the difference in the signal transduction pathway leading to the cellular responses at high and low concentrations; i.e., the differentiative effect of PGE₂ is mediated by cyclic adenosine monophosphate (cAMP), while the proliferative effect is mediated through the elevation of intracellular Ca²⁺ concentration ([Ca²⁺]_i). By contrast, we have recently shown, using adult rat calvarial cells, that PGE₂ stimulates bone nodule (BN) formation via the Ca²⁺ pathway at low and high concentrations [Kaneki et al., 1999]. The results indicate that the transduction mechanism utilized for PGE₂ signal may differ depending on the cell type. The mode of response to PGE₂ is known to be affected also by factors other than PGE₂ concentration. When cell fractions obtained after serial enzyme digestions of fetal parietal bone were treated with PGE₂, DNA synthesis was increased in the earliest fraction used, while it was decreased in a mixture of cells from the last three fractions [Centrella et al., 1994]. It has also been shown that the time at which PGE₂ is added during the culture period to MC3T3-E1 cells affects the responses to the agonist [Suda et al., 1996]. The diversity of the osteoblastic cell responses to PGE₂ may reflect the difference in the stage of differentiation; i.e., the signal transduction mechanism underlying the responses to PGE₂ may be modulated during the differentiation process. Furthermore, the *in vivo* effect of PGE₂ on bone metabolism in rats has been reported to be age dependent [Jee et al., 1985, 1990]. Within this context, it is interesting to determine the cell donor age-related change in the effect of PGE₂ on osteoblastic cells.

The purpose of this study was to examine the change in the ability of the rat calvarial osteoblast-like cells to proliferate and differentiate, to form mineralized BN with age of the cell donor. Age-related changes in the signal transduction mechanism were also studied.

MATERIALS AND METHODS

Materials

The following prostaglandin analogues were purchased from Cayman Chemical (Ann Arbor, MI): 17-phenyl- ω -trilor PGE₂ (EP₁ agonist) and 11-deoxy-PGE₁ (EP₂/EP₄ agonist). PGE₂, forskolin, and 3-(isobutyl)-1-methylxanthine (IBMX) were from Sigma Chemical Co. (St. Louis, MO).

Fetal bovine serum (FBS), phenol red-free F-12 medium, and α -minimum essential medium (α -MEM), and trypsin were products of Gibco-BRL (Rockville, MD). Kanamycin, ascorbic acid, collagenase, and β -glycerophosphate were obtained from Wako Pure Chemical Industries (Tokyo, Japan). [Methyl-³H]thymidine was from DuPont (Wilmington, DE).

Cell Culture

Cells enriched for osteoblast phenotype were enzymatically isolated from the calvaria of 4- to 120-week-old female Wistar rats (CLEA, Tokyo, Japan) according to the method of Bellows et al. [1986], modified by Kato et al. [1995]. Briefly, after five sequential digestions of calvaria with a mixture of collagenase and trypsin, the released cells from the last three digestion intervals were grown in F-12 medium containing 10% FBS. After reaching confluence, the cells were collected by a trypsin treatment, seeded in 4-well dishes at 2×10^3 cells/cm² in the same medium, and cultured for 4 days (designated as the proliferation period, P1-P4). At the end of day P4, the medium was changed to α -MEM supplemented with 10% FBS, 2 mM β -glycerophosphate and ascorbic acid (0.1 mg/ml), and the cells were maintained for a further 18 days (designated as the mineralization period, M1-M18).

Determination of Markers for Cell Proliferation and Differentiation

For the determination of alkaline phosphatase (ALP) activity, BN formation, and DNA synthesis, cells were incubated in serum-free F-12 medium for 24 h on day P4, with 10^{-6} M PGE₂ present in the medium as during the last 6 h; the incubation was then continued in PGE₂-free α -MEM medium, as described above. An ALP assay was performed at the beginning of day M2 according to the method of Lowry et al. [1954], using p-nitrophenylphosphate as a substrate. Quantification of BN was performed by visualization with von Kossa stain [Bhargava et al., 1988], and the total nodule area and the number of nodules were assessed on day M18, using a colony counter (BMS-400, Toyo Sokki, Tokyo, Japan). For determination of DNA synthesis, the cells were incubated for 3 h in serum-free α -MEM medium containing [³H]thymidine (1.25 μ Ci/ml) at the beginning of day M1, and incorporation of radioactivity into DNA was measured.

Determination of cAMP and Inositol Triphosphate Formation

Intracellular cAMP was determined using a radioimmunoassay (RIA) kit (Yamasa, Chiba, Japan) as previously described [Kaneki et al. 1999]. Determination of the inositol 1,4,5-triphosphate (IP₃) generation was performed using an RIA kit (Amersham, Tokyo, Japan) as described [Kaneki et al., 1999].

Determination of PGE₂ Production

Calvarial cells were cultured for various periods under standard conditions; serum was then omitted from the medium for 18 h. The medium was replaced with fresh serum-free medium with or without NS-398 (10⁻⁶ M), and the incubation was continued for 3 h. The medium was changed again to serum-free standard medium, and the culture medium was collected after an additional 3-h incubation. The amount of PGE₂ was measured using the Prostaglandin E₂ [¹²⁵I] Assay System (Amersham Pharmacia Biotech, Tokyo, Japan).

Statistical Methods

Data were analyzed by Student's *t*-test or by one-way analysis of variance (ANOVA), followed by Dunnett's multiple comparison test. *P* < 0.01 was considered significant. All data are presented as the mean ±SD of four cultures. The experiments with rats aged <100 weeks were repeated at least three times, while those performed with rats aged ≥100 weeks were repeated twice.

RESULTS

First, we examined the age-dependent changes in the parameters for proliferation and differentiation in cells not treated with PGE₂ (Fig. 1). BN formation in the area (Fig. 1A), number (Fig. 1B), and activity of ALP (Fig. 1C), continuously decreased with the age of the cell donor at 6–50 weeks and then remained at a relatively constant level at ≤120 weeks. The profiles of BN formed by the cells from 6- and 90-week-old rats are shown in Figure 2. A similar age-dependent decline was observed in the rate of [³H]thymidine incorporation into the cells (Fig. 1D). These results suggest that the spontaneous proliferation and differentiation of the calvarial cells are suppressed as a function of donor age at 6–50 weeks.

To determine the role of endogenous PGE₂, we examined the change in the PGE₂ production during the culture of the cells from 10-, 25-, and 90-week-old rats (Fig. 3). Sharp peaks of PGE₂ production were obtained on day P4, regardless of the cell donor age. The addition of a cyclooxygenase-2 (COX-2) inhibitor, NS-398 (10⁻⁶ M), to the culture almost completely blocked PGE₂ production (Fig. 3), indicating that PGE₂ is generated exclusively through a COX-2 pathway. We therefore treated calvarial cells from rats of various ages with NS-398 on day P4 (Fig. 4). NS-398 decreased the area of BN by approximately 50% (Fig. 4) in the cells from 10- to 35-week-old rats, indicating that the endogenous PGE₂ increases the area of BN. By contrast, the area of BN was increased by NS-398 by approximately 20–40% (Fig. 4) in the cells from 90- to 120-week-old rats. These results suggest that the response of calvarial cells to PGE₂ differs depending on the age of the cell donor.

We next determined the effects of exogenous PGE₂ on proliferation and differentiation in cells from rats of various ages (Fig. 5). As expected from the results shown in Figure 3, the BN formation was enhanced by the PGE₂ treatment, with respect to both area (Fig. 5A) and number (Fig. 5B) by 100–180% and by 50–90%, respectively, in the cells from 4- to 35-week-old rats. Rats in this age range are hereafter referred to as young rats. The PGE₂ treatment also enhanced the ALP activity by 60–130% in the cells from young rats (Fig. 5C). The maximal percentage stimulation of BN formation and ALP activity by PGE₂ was observed in the cells from 10-week-old rats; the effect was gradually decreased with age. PGE₂ significantly decreased the area of BN (Fig. 5A) and ALP activity (Fig. 5C) in the cells from 80- to 120-week-old rats. Rats in this age range are hereafter referred to as aged rats. By contrast, PGE₂ both suppressed and enhanced [³H]thymidine incorporation into the cells from young rats and aged rats, respectively (Fig. 5D). An agonist specific for the EP₁ receptor subtype of PGE₂, 17-phenyl-ω-trinor PGE₂, showed almost the same effects as PGE₂ on the proliferation and differentiation markers in the cells from young rats (Fig. 6). It increased the BN formation in both area (Fig. 6A) and number (Fig. 6B) and the ALP activity (Fig. 6C) by 130–190%, 60–100%, and 110–140%, respectively. Maximal percentage stimulation was observed in the

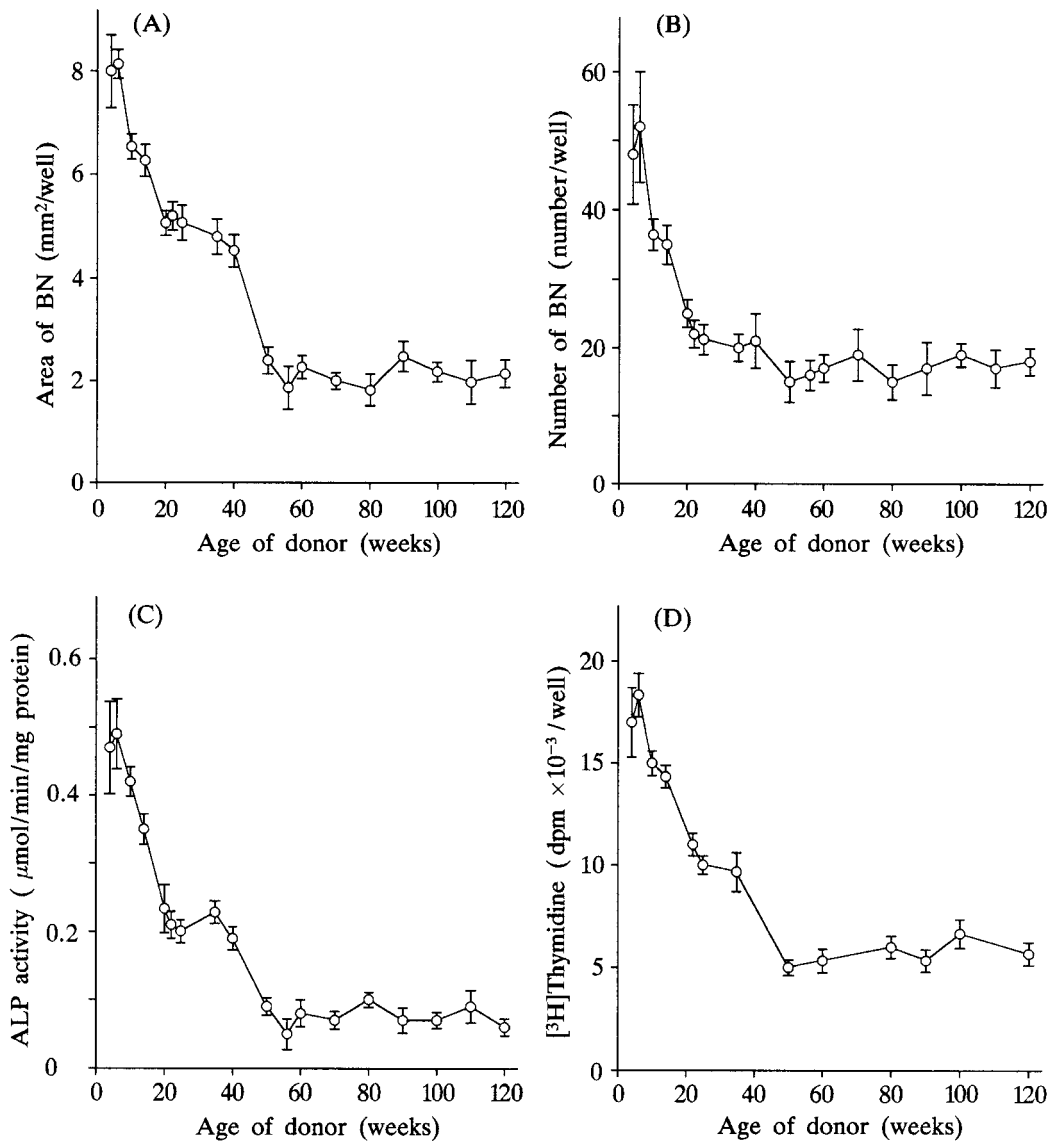


Fig. 1. Age-related changes in bone nodule (BN) formation, alkaline phosphatase (ALP) activity, and DNA synthesis in cultures of calvarial cells from rats. ALP activity (C) was measured at the beginning of day M2. [³H]Thymidine incorporation into the cells (D) was measured at the beginning of day M1. The area (A) and number (B) of BN were measured on day M18. Other conditions were as described under Materials and Methods. Each value represents the mean \pm SD of four cultures. The experiments with rats aged <100 weeks were repeated at least three times, while those with rats aged ≥ 100 weeks were repeated twice, and the results were essentially the same as those depicted.

cells from 10- to 15-week-old rats and the effects were gradually reduced with age. The EP₁ agonist decreased [³H]thymidine incorporation into the cells by 20–50% in the cells from young rats (Fig. 6D). By contrast, with PGE₂, the EP₁ agonist showed no significant effect on the BN formation, ALP activity and [³H]thymidine incorporation into the cells from aged rats. These results suggest that the effects of PGE₂ on both proliferation and differentiation are exerted ex-

clusively through the EP₁ receptor subtype in the cells from young rats and that the signal transduction pathway through EP₁ is not active in the cells from aged rats.

A selective agonist for EP₂ and EP₄ receptor subtypes, 11-deoxy-PGE₁, decreased the BN formation in both area (Fig. 7A) and number (Fig. 7B) and the ALP activity (Fig. 7C) by approximately 50%, 20%, and 50%, respectively, at all cell donor ages. By contrast, 11-deoxy-PGE₁

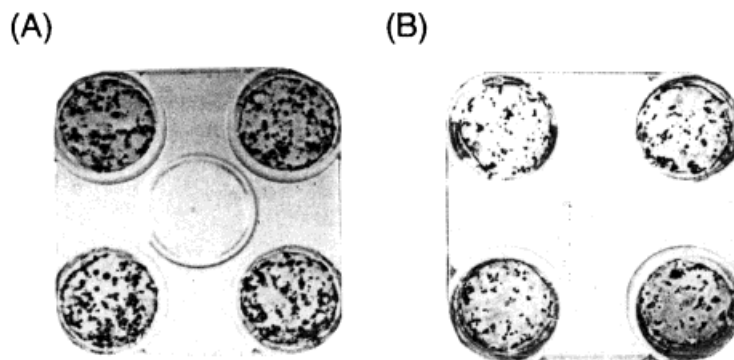


Fig. 2. Phase-contrast photomicrographs of bone nodules (BN) visualized with von Kossa stain in cultures of calvarial cells from 6-week-old rats (A) and 90-week-old rats (B).

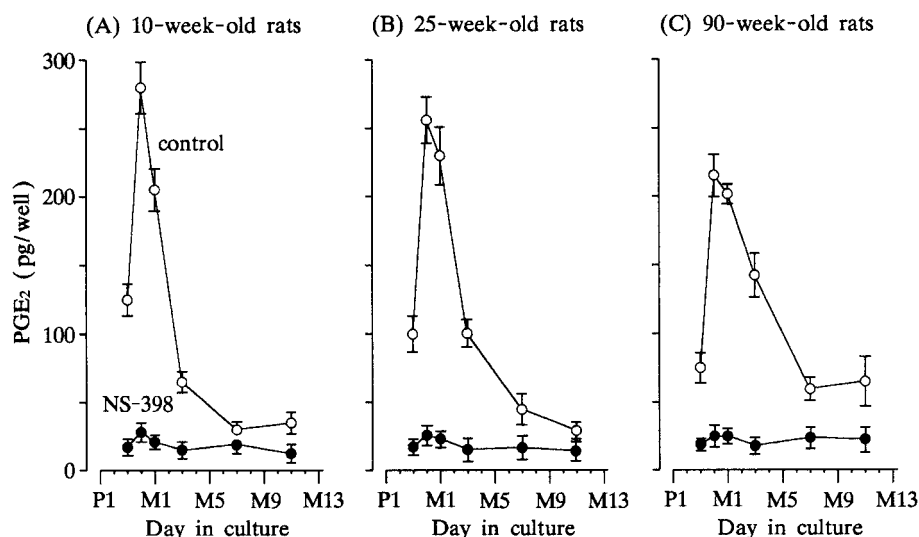


Fig. 3. Time-course of prostaglandin E₂ (PGE₂) production in the cultures of calvarial cells from 10-week-old rats (A), 25-week-old rats (B), and 90-week-old rats (C). The culture medium was collected on the indicated days and PGE₂ in the medium was determined as described under Materials and Methods. After preincubation with (●) or without (○) NS-398 (10⁻⁶ M) and a

further 3-h incubation in the serum-free standard medium, endogenous PGE₂ was determined on the indicated days. Each value represents the mean \pm SD of four cultures. Experiments were repeated at least three times, and results were essentially the same as those depicted.

enhanced the rate of [³H]thymidine incorporation into the cells by approximately 70% at all cell donor ages examined (Fig. 7D). These results indicate that activation of the EP₂/EP₄ pathway leads to the suppression of the cell differentiation and the enhancement of the cell proliferation, regardless of the cell donor age. The activation of EP₂/EP₄ pathway is known to cause an increase in the activity of adenylate cyclase to produce cAMP [Suda et al., 1996]. We therefore tested the effect of a direct activator of adenylate cyclase, forskolin, on the markers for differentiation and proliferation of osteoblastic cells (data not shown); the results with forskolin are essentially the same as those with the EP₂/EP₄ agonist shown in Figure 7. These ef-

fects of forskolin closely matched those of 11-deoxy-PGE₁, indicating that the EP₂/EP₄ agonist exerts its effects on osteoblastic cells by the activation of adenylate cyclase and the subsequent production of cAMP.

We then compared the effects of PGE₂, 17-phenyl- ω -trilor PGE₂, 11-deoxy-PGE₁ and forskolin on the production of cAMP (Fig. 8A) and IP₃ (Fig. 8B) in the cells from 25- and 90-week-old rats. PGE₂, 11-deoxy-PGE₁, and forskolin, but not 17-phenyl- ω -trilor PGE₂, induced the production of cAMP in the cells from the rats of both ages. By contrast, PGE₂ and 17-phenyl- ω -trilor PGE₂ enhanced the production of IP₃ in the cells from 25-week-old rats, but not in the cells from 90-week-old rats. These results indi-

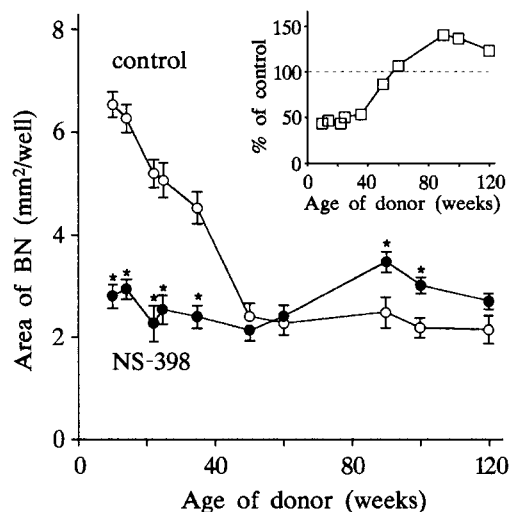


Fig. 4. Effect of the inhibition of endogenous prostaglandin E₂ (PGE₂) production by NS-398 on the area of bone nodules (BN) in cultures of calvarial cells from rats of various ages. The cells were treated with (●) or without (○) NS-398 (10⁻⁶ M) for 3 h on day P4, and the area of BN was measured on day M18. Inset compares the effect of NS-398 on the ratio of the area of BN in NS-398 treated cultures with that in untreated cultures. Other conditions were as described under Materials and Methods. Each value represents the mean ±SD of four cultures. The experiments with rats aged <100 weeks were repeated at least three times, while those with rats aged ≥100 weeks were repeated twice, and the results were essentially the same as those depicted.

cate that the signal transduction pathway through EP₁ receptor subtype is inactive in the cells from 90-week-old rats.

DISCUSSION

To the best of our knowledge, this is the first report to show a donor age-dependent change in the responsiveness of osteoblast-like cells to PGE₂ in primary culture. These age-dependent changes in the properties of the cells could at least partially be explained by the alteration of the signal transduction mechanisms for endogenously generated PGE₂, because the BN formation is largely affected by pretreatment of the cells with a COX-2 inhibitor; only a minor change with aging of the cell donor was observed in the rate of endogenous PGE₂ production. PGE₂ receptors are classified into four subtypes, EP₁, EP₂, EP₃, and EP₄ [Coleman et al., 1994]. Experiments using agonists specific for the PGE₂ receptor subtypes demonstrated the presence of EP₁ and EP₂/EP₄ in MC3T3-E1 osteoblast-like cells [Suda et al., 1996]. EP₁ couples to the phosphoinositide/Ca²⁺ pathway, while EP₂/EP₄ couple to the stimulative adeny-

late cyclase/cAMP pathway [Coleman et al., 1994]. Using adult rat (25- to 35-week-old) calvarial cells, we previously showed that PGE₂ stimulates the increase in the concentration of intracellular Ca²⁺ ([Ca²⁺]_i) and production of cAMP through EP₁ and EP₂/EP₄ receptor subtypes, respectively [Kaneki et al., 1999], as illustrated in Figure 9. The increase in [Ca²⁺]_i leads to the activation of ALP, collagen synthesis, and BN formation, as well as suppression of cell proliferation. Although cAMP has the ability to suppress ALP activity, collagen synthesis, and BN formation and to enhance proliferation, these effects of cAMP are not observable because the newly formed cAMP is rapidly degraded by phosphodiesterase, which is activated by the increase in [Ca²⁺]_i [Kaneki et al., 1999].

In the present study, PGE₂ and an EP₁ agonist, 17-phenyl- ω -trilor PGE₂, enhanced the BN formation and ALP activity and suppressed proliferation of the cells from young rats. By contrast, an adenylate cyclase activator, forskolin, and an EP₂/EP₄ agonist, 11-deoxy-PGE₁, suppressed the BN formation and ALP activity and enhanced proliferation of these cells. These results suggest that the effect of PGE₂ on the cells from young rats is mainly exerted through the EP₁ receptor subtype (Fig. 9). In the cells from aged rats, PGE₂, forskolin, and 11-deoxy-PGE₁ significantly decreased the BN formation and ALP activity, while 17-phenyl- ω -trilor PGE₂ showed no effect on these differentiation markers. By contrast, cell proliferation was enhanced by PGE₂, forskolin and 11-deoxy-PGE₁ and not affected by 17-phenyl- ω -trilor PGE₂. These results can be explained by assuming that the signal transduction pathway through EP₁ is inactive in the cells from aged rats and PGE₂ suppresses the cell differentiation and enhances the cell proliferation through the EP₂/EP₄-cAMP pathway (Fig. 9). If the EP₁ pathway is inactive, phosphodiesterase, which degrades newly formed cAMP, is not activated; therefore, cAMP may accumulate, suppressing differentiation and enhancing proliferation of the cells.

The cellular responses through the EP₁ and EP₂/EP₄ pathway in rat calvarial cells are different from those observed with MC3T3-E1 cells in which cAMP stimulates cell differentiation and the activation of PKC results in the stimulation of proliferation and the suppression of differentiation [Hakeda et al., 1986, 1987]. Furthermore, Suda et al. [1996] have reported the presence of EP₁ and EP₂/EP₄ subtypes of PGE₂

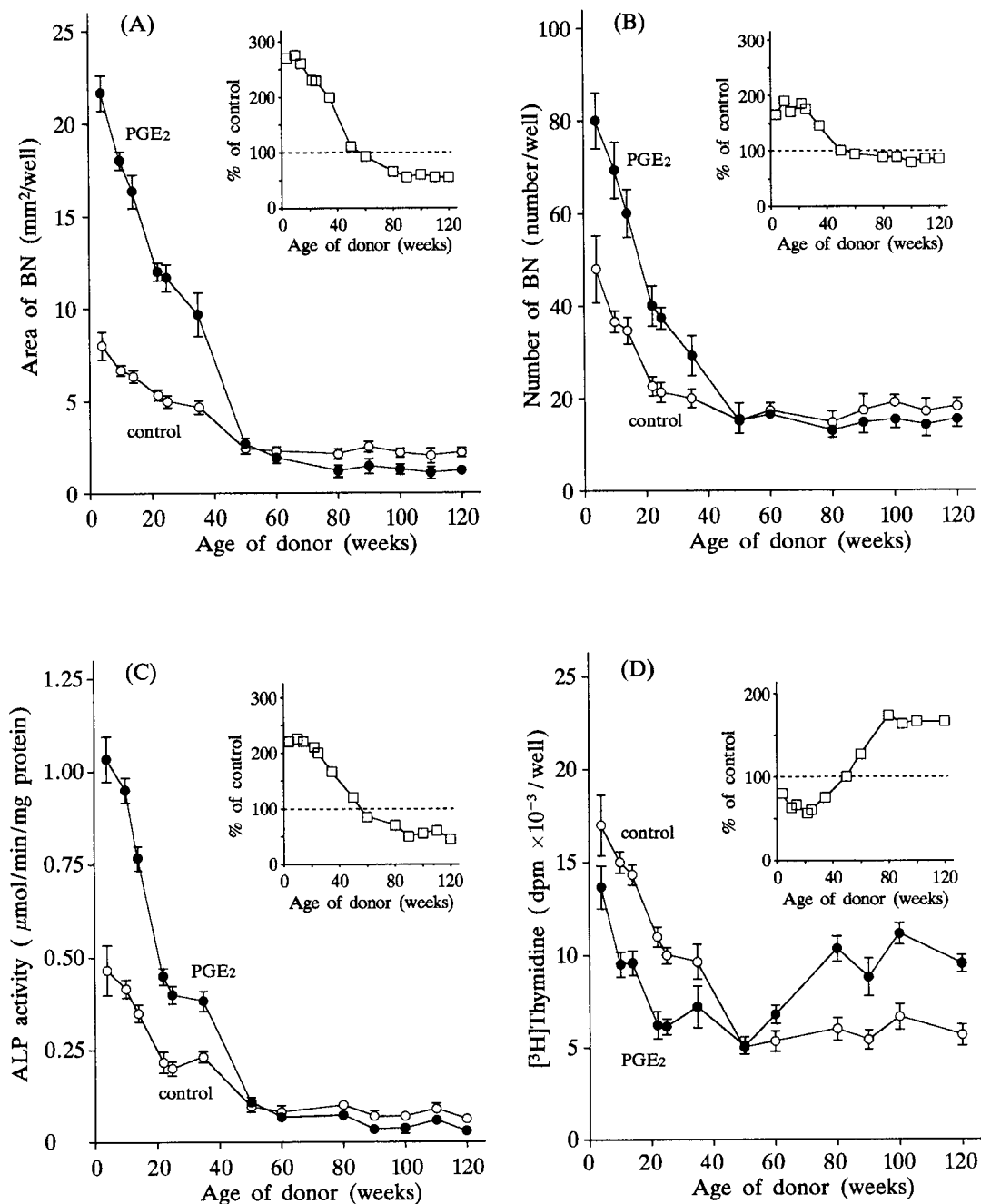


Fig. 5. Effects of prostaglandin E₂ (PGE₂) on bone nodule (BN) formation, alkaline phosphatase (ALP) activity, and DNA synthesis in cultures of calvarial cells from rats of various ages. The cells were treated with (●) or without (○) PGE₂ (10⁻⁶ M) during the last 6 h of day P4. ALP activity (C) was measured at the beginning of day M2. [³H]Thymidine incorporation into the cells (D) was measured at the beginning of day M1. The area (A) and number (B) of BN were measured on day M18. Inset

compares the effect of PGE₂ on the ratio of the BN formation, ALP activity, and [³H]thymidine incorporation in PGE₂-treated cultures with that in untreated cultures. Other conditions were as described under Materials and Methods. Each value represents the mean ±SD of four cultures. The experiments with rats aged <100 weeks were repeated at least three times, while those with rats aged ≥100 weeks were repeated twice, and the results were essentially the same as those depicted.

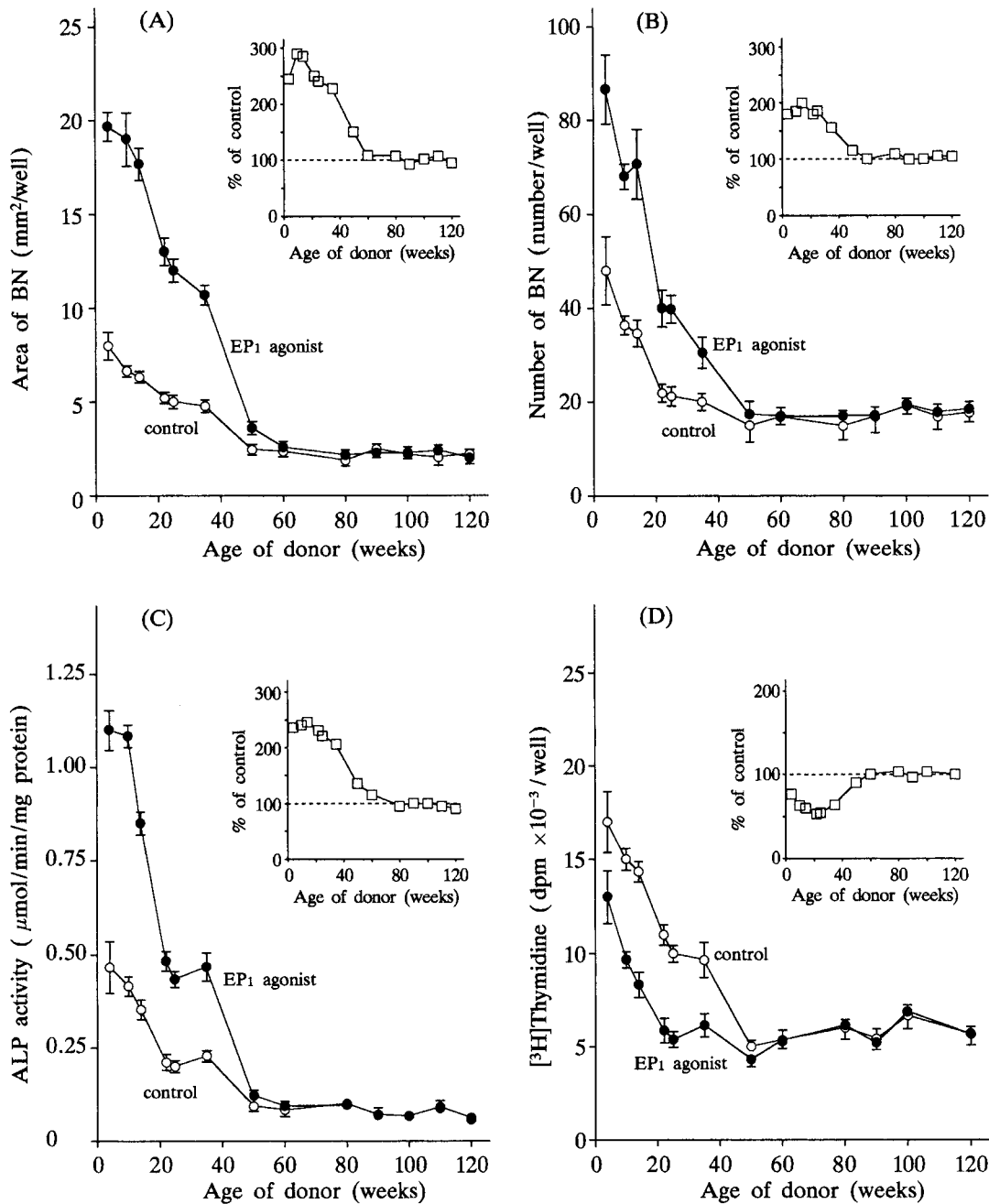


Fig. 6. Effects of EP₁ agonist, 17-phenyl- ω -trinor prostaglandin E₂ (PGE₂), on the bone nodule (BN) formation, alkaline phosphatase (ALP) activity, and DNA synthesis in cultures of calvarial cells from rats of various ages. The cells were treated with (●) or without (○) 17-phenyl- ω -trinor PGE₂ (10⁻⁶ M) during the last 6 h of day P4. ALP activity (C) was measured at the beginning of day M2. [³H]Thymidine incorporation into the cells (D) was measured at the beginning of day M1. The area (A) and number (B) of BN were measured on day M18. Inset compares the effect of

17-phenyl- ω -trinor PGE₂ on the ratio of the BN formation, ALP activity, and [³H]thymidine incorporation in 17-phenyl- ω -trinor PGE₂-treated cultures with that in untreated cultures. Other conditions were as described under Materials and Methods. Each value represents the mean \pm SD of four cultures. The experiments with rats aged <100 weeks were repeated at least three times, while those with rats aged \geq 100 weeks were repeated twice, and the results were essentially the same as those depicted.

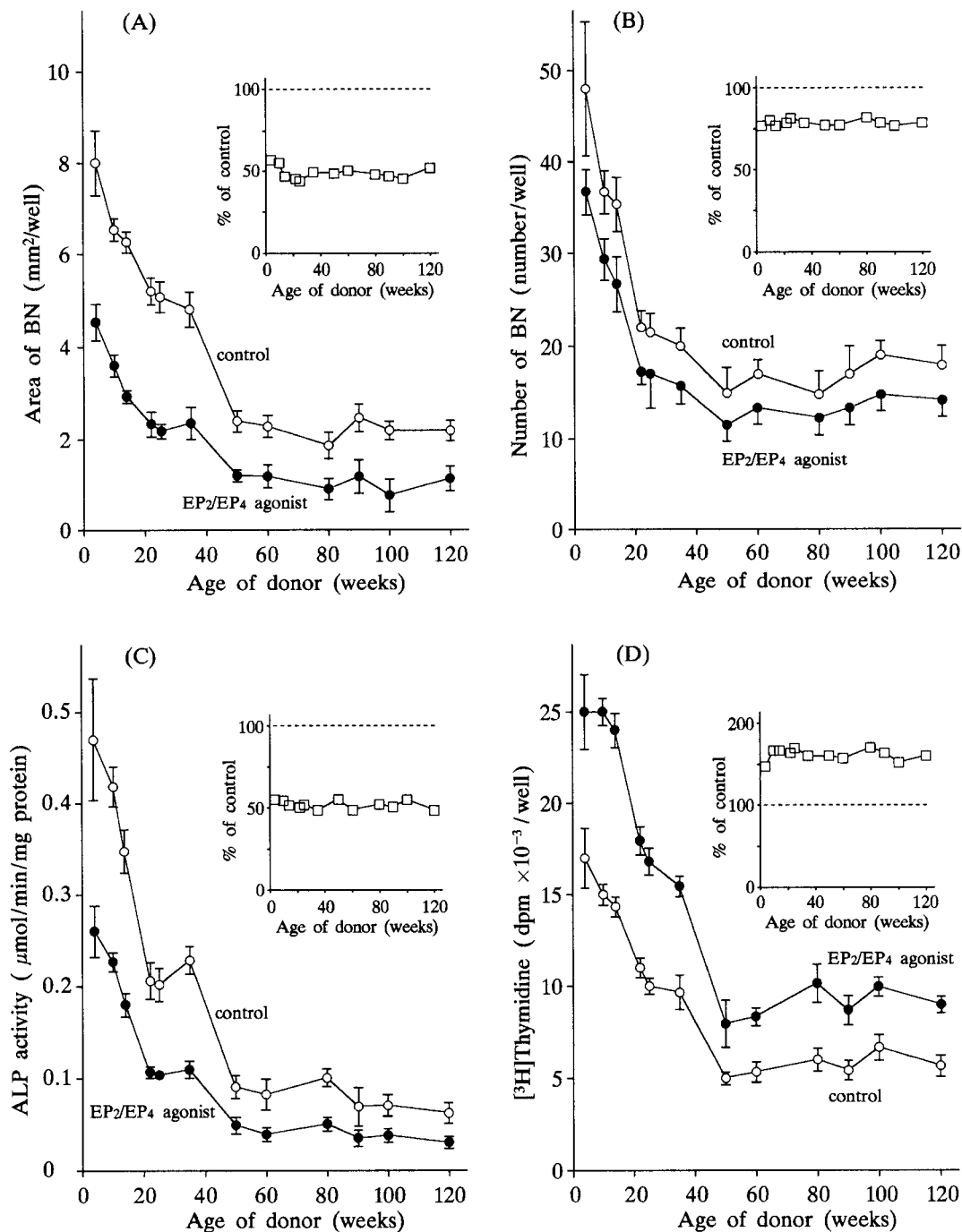


Fig. 7. Effects of EP₂/EP₄ agonist, 11-deoxy-prostaglandin E₁ (PGE₁), on bone nodule (BN) formation, alkaline phosphatase (ALP) activity, and DNA synthesis in cultures of calvarial cells from rats of various ages. The cells were treated with (●) or without (○) 11-deoxy-PGE₁ (10⁻⁶ M) during the last 6 h of day P4. ALP activity (C) was measured at the beginning of day M2. [³H]Thymidine incorporation into the cells (D) was measured at the beginning of day M1. The area (A) and number (B) of BN were measured on day M18. Inset compares the effect of

11-deoxy-PGE₁ on the ratio of the BN formation, ALP activity, and [³H]thymidine incorporation in 11-deoxy-PGE₁-treated cultures with that in untreated cultures. Other conditions were as described under Materials and Methods. Each value represents the mean ± SD of four cultures. The experiments with rats aged <100 weeks were repeated at least three times, while those with rats aged ≥100 weeks were repeated twice, and the results were essentially the same as those depicted.

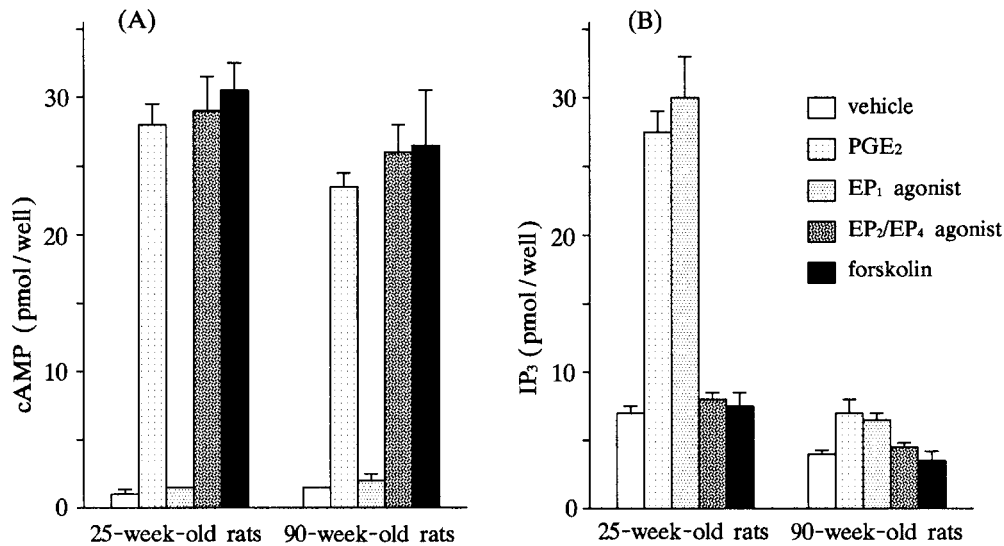


Fig. 8. Effects of prostaglandin E₂ (PGE₂), the agonists specific for PGE₂ receptor subtypes, and forskolin on the accumulation of cyclic adenosine monophosphate (cAMP) and IP₃ generation in the cultures of calvarial cells from 25-week-old rats (A) and 90-week-old rats (B). For the determination of cAMP, the cells were incubated in serum-free F-12 medium for 18 h at the beginning of day P4 and treated with 10⁻⁵ M 3-(isobutyl)-1-methylxanthine (IBMX) for 20 min, followed by 10⁻⁶ M PGE₂, 10⁻⁶ M 17-phenyl- ω -trior PGE₂ (EP₁ agonist), 10⁻⁶ M

PGE₁ (EP₂/EP₄ agonist) or 10⁻⁶ M forskolin for 5 min. For the determination of IP₃, the cells were incubated in inositol- and serum-free medium containing 0.01% bovine serum albumin for 18 h at the beginning of day P4. The cells were then treated with the four agonists (10⁻⁶ M) for 0.5 min. Other conditions were as described under Materials and Methods. Each value represents the mean \pm SD of four cultures. The experiments were repeated at least three times, and the results were essentially the same as those depicted.

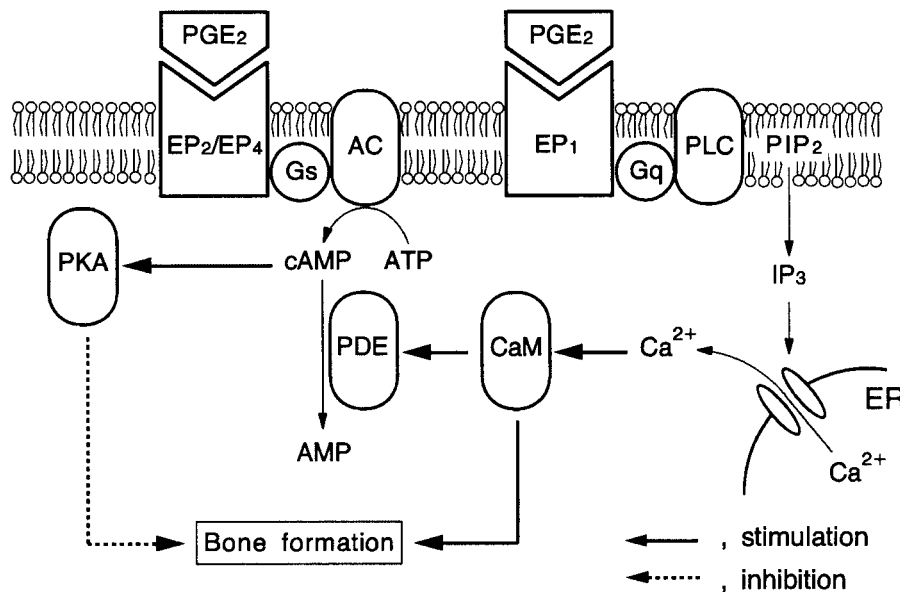


Fig. 9. Scheme of the signal transduction pathway for prostaglandin E₂ (PGE₂)-induced cellular responses in rat calvarial cells. Binding of PGE₂ to the EP₂/EP₄ receptor subtype leads to the activation of adenylate cyclase (AC) to produce cyclic adenosine monophosphate (cAMP), which inhibits bone formation through the activation of cAMP-dependent protein kinase (PKA). Binding of PGE₂ to the EP₁ receptor subtype leads to the

activation of phospholipase C (PLC) to produce IP₃, which binds to its receptor on endoplasmic reticulum (ER), resulting in the release of Ca²⁺. The increase in [Ca²⁺]_i causes the activation of bone formation. Ca²⁺ also activates a phosphodiesterase (PDE) that blocks the signaling through EP₂/EP₄ receptor subtype by degrading cAMP.

receptor and proposed that the former promotes cell growth, whereas the latter mediates differentiation of osteoblasts. In the present study, the EP₁ agonist promoted differentiation and suppressed growth, while the EP₂/EP₄ agonist promoted growth and suppressed differentiation. As described in a previous paper [Kaneki et al., 1999], the increase in [Ca²⁺]_i, but not the activation of PKC transduces EP₁ signal in rat calvarial cells. The reason for the discrepancy remains unknown, but one possible explanation would be the difference in the stage of differentiation. It has been reported that PGE₂ treatment increases DNA synthesis in the first fraction of five serial collagenase digestion, while it inhibits DNA synthesis in the mixture of third to fifth fractions [Centrella et al., 1994]. The results were explained by the difference in the differentiation stage between the two cell populations. The mode of the response to PGE₂ is also affected by the time of addition during the culture period [Suda et al., 1996; Kaneki et al., 1999]. These differences in the cellular responses to PGE₂ may be a reflection of the difference in the differentiation stage, because osteoblastic cells are likely to differentiate spontaneously during the culture. The present study suggested that the EP₁ pathway is not functioning in the aged rats. The lack of signaling through EP₁ receptor subtype may be attributable to the donor age-dependent loss of the receptor subtype. The changes in the expression of the receptor subtypes with age remain to be examined.

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